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3 **Doxycycline levels and anti-Wolbachia antibodies in sera from dogs experimentally
4 infected with *Dirofilaria immitis* and treated with a combination of
5 ivermectin/doxycycline**

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18

19 **Abstract**

20 Sera from *Dirofilaria immitis*-experimentally infected dogs treated with a combination of
21 ivermectin/doxycycline were analysed for doxycycline levels by HPLC and anti-
22 *Wolbachia* Surface Protein (rWSP) antibodies by ELISA and compared with sera from
23 dogs treated with doxycycline alone. Results show that doxycycline levels were not
24 statistically different between the two groups. Circulating anti-WSP antibody titres were
25 markedly lower in both treatment groups when compared to control *D. immitis* infected
26 dogs, indicating that doxycycline is able to reduce *Wolbachia* and prevent the immune
27 response against the bacteria. The combination treatment protocol has been shown to be
28 highly adulticidal and further studies are needed to better understand the interaction
29 between doxycycline and ivermectin in *D. immitis* infected dogs.

30

31 **1. Introduction**

32 Heartworm infection (HW; *Dirofilaria immitis*) in dogs causes chronic pulmonary disease
33 that, if left untreated, can lead to right-side congestive heart failure. Currently, the only
34 registered drug for adulticide therapy in dogs with heartworm disease (HWD) is
35 melarsomine dihydrochloride (Immiticide®, Merial). Due to concerns of severe, post-
36 treatment thromboembolism in some dogs (Kramer et al., 2008) and recent problems with
37 availability of melarsomine on several international markets, there is increasing interest in
38 alternative adulticide treatments (Colby et al., 2011).

39 The recent targeting of the bacterial endosymbiont *Wolbachia*, through antibiotic therapy
40 of the infected host, has offered an interesting alternative for the treatment of HWD.

41 Indeed, *Wolbachia* is necessary for the reproductive capacity and long-term survival of
42 those filarial parasites that harbour the endosymbiont. The adulticide effects of
43 doxycycline (DOXY) have been studied in *D. immitis* experimentally infected dogs
44 (Bazzocchi et al., 2008). No significant adulticide effects at 8 months post infection
45 following several cycles of DOXY was observed, even though treatment was able to reduce
46 *Wolbachia* populations. The same study reported that when DOXY was combined with the
47 macrocyclic lactone ivermectin (IVM), adulticide efficacy was approximately 80% vs. 9%
48 when dogs were treated with DOXY alone. The adulticide effect of this combination has
49 also been confirmed in naturally infected dogs (Grandi et al., 2010). It is not clear why the
50 two drugs work better together in eliminating a large population of heartworms in a
51 relatively short period of time (8–10 months).

52 drugs, including in pharmacokinetics. The present study was aimed at evaluating DOXY
53 levels and circulating antibodies against *Wolbachia* Surface Protein (WSP) in serum from
54 dogs treated with DOXY alone or in combination with IVM, according to Bazzocchi et al.
55 (2008).

56

57 **2. Materials and Methods**

58 **2.1. Animals and sera**

59 Briefly, serum samples conserved at -20°C from a previous study of *D. immitis*-
60 experimentally infected dogs were used (Bazzocchi et al., 2008). Treatment protocols are
61 reported in Table 1. Each group consisted of five dogs experimentally infected with adult
62 heartworms (7 males and 9 females) by intravenous transplantation. Drugs were given with

63 food in the morning and samples were taken at approximately 6 h later. Serum samples
64 from 2 drug administration days, corresponding to the weekly IVM treatment, were
65 analysed for drug concentrations: T1 (6 weeks postinfection, p.i.) and T4 (34 weeks p.i.).
66 For anti-WSP ELISA, serum samples from T0 (6 weeks before infection to determine cut-
67 off values), T1, T2 (10 weeks p.i.) and T4 were analysed.

68

69 2.2. HPLC for doxycycline serum levels

70 The concentrations of DOXY in serum were measured by means of HPLC method,
71 following the technique by Nielsen and Gyrd-Hansen (1996), slightly modified. The HPLC
72 system consisted of a Prostar LC Workstation (Varian Co., Walnut Creek, CA, USA), with
73 a Prostar 325 UV-Vis detector and a 10 L loop. Chromatographic separations were
74 obtained using a Syncronis C18 analytical column (Thermo, Milan, Italy) (5 m particle
75 size, 150 mm × 4.6 mm), maintained at room temperature (20°C). The analytical
76 wavelength was set at 350 nm. The mobile phase consisted of acetonitrile and 0.01 mol/L
77 trifluoroacetic acid (30:70, v/v), with a flow rate of 1.0 mL/min. All used solvents and
78 reagents were of HPLC grade purity and were purchased from Sigma–Aldrich
79 (Milan, Italy).

80 Samples were prepared by adding 400 L buffer EDTA (0.1 mol/L sodium phosphate,
81 containing 0.1 mol/L disodium EDTA; pH of the buffer mixture was adjusted to 5.0 by
82 adding 0.1 mol/L phosphoric acid) and 100 L perchloric acid 20% to 500 L of serum and
83 the mixture was placed in vortex mixer for 2 min and then centrifuged at 7500 × g for 20
84 min. The supernatant was collected, filtered through a

85 0.22-m syringe filter, put in sample vial and injected into the HPLC system. A serum
86 sample from a *D. immitis* infected dog receiving no treatment was used as negative control.

87

88 2.3. ELISA for anti-WSP antibodies

89 The recombinant protein WSP of the *Wolbachia* of *D. immitis* (rWSP) was produced in
90 *Escherichia coli* and purified as described in Bazzocchi et al. (2000). Wells of ELISA flat-
91 bottom plates were coated with 0.1 g/well of rWSP. Sera were analysed in duplicate at a
92 dilution of 1:100 and the anti-dog IgG HRP-conjugated antibody (Sigma–Aldrich) was
93 diluted at 1:5000. The optical density (O.D.) was measured at 492 nm. The cut-off was
94 established at an O.D. of 0.65, which is the mean O.D. of the control sera (sera from each
95 dog at the moment of infection) plus three times their standard deviation. Samples with
96 O.D. less than of 0.65 were classified as negative and samples with O.D. greater than or
97 equal to 0.65 were classified as positive.

98

99 2.4. Statistical analysis

100 Differences in DOXY serum levels (mg/L) at each time point were analysed by comparing
101 median values by Mann–Whitney U test (Genstat, 7th edition) and $p < 0.05$ was considered
102 to be a significant difference.

103

104 **3. Results and discussion**

105 Serum levels of antibiotic in dogs treated with the combination IVM/DOXY protocol were
106 not statistically different compared to dogs treated with DOXY alone at any time points

107 considered (Fig. 1). Therefore it is unlikely that the adulticide effect of the combination
108 treatment shown in the previous study was due to a difference in tissue/worm distribution
109 of DOXY. There was, however, a wide range of variability in serum concentrations among
110 dogs and among time points, making interpretation of results difficult. Interestingly, dogs
111 from both the combination group and the DOXY group showed markedly lower values for
112 anti-
113 WSP antibodies when compared to untreated HW-infected controls (Fig. 2). This is
114 strongly suggestive of elimination of *Wolbachia* from *D. immitis*, as previously shown by
115 PCR analysis of worms collected from treated dogs at necropsy (Bazzocchi et al., 2008).
116 So, DOXY, whether alone or in combination, is actively eliminating *Wolbachia* from adult
117 worms efficiently enough to prevent the antibody response to it. Yet, this is not sufficient
118 for an adulticide effect greater than 9% (Bazzocchi et al., 2008). Only the combination of
119 DOXY with IVM is able to kill the parasite. If the antibiotic is taken up and distributed in
120 a uniform way in both protocols, and the effect on *Wolbachia* is comparable, it may be that
121 the interaction in the combination protocol is synergistic. Indeed, it is possible that DOXY
122 has a detrimental effect on *D. immitis* independent of its effect on *Wolbachia*, as has been
123 suggested previously (Smith and Rajan, 2000). IVM causes neuromuscular dysfunction,
124 pharyngeal paralysis, and thickening of the gut epithelium in treated worms. Ultrastructural
125 analysis of IVM-treated *D. immitis* show retained ingesta and increased gut permeability
126 (Steffens and McCall, 1998). These alterations may lead to an increase in the concentration
127 of DOXY within the worm. The two drugs may also be interacting on a molecular level: it
128 has been reported that IVM is able to reduce cellular efflux of antibiotics in farm animals,

129 thus increasing the intracellular concentration of the latter. It would appear that this is due
130 to IVM's ability to inhibit the activity of various cellular transport systems (Lespine et al.,
131 2006; Real et al., 2011; Ballent et al., 2012). On the other hand, it cannot be excluded that
132 DOXY in some way potentiates the effects of IVM, even though this seems less likely.
133 However, several compounds, including antibiotics, have been shown to increase
134 intracellular concentrations of macrocyclic lactones such as moxidectin (Dupuy et al.,
135 2006).

136 Furthermore, since tetracycline was shown to inhibit oxidation of fatty acids in
137 mitochondria of mice and man (Fréneaux et al., 1988), it is possible that DOXY could
138 exert a toxic effect also on the nematode mitocondria. Finally, tetracyclines are known to
139 bind to bivalent ions such as calcium and magnesium. An intriguing hypothesis could be
140 that DOXY may interfere with calcium uptake into parasite neurons. Indeed, it was
141 observed that minocycline is able to cause calcium-dependent neuromuscular block in
142 rabbits (Hashimoto et al., 1979). It is therefore possible that partial neuromuscular block
143 induced by doxycycline could add to the paralyzing effect by IVM in a synergistic fashion,
144 thus resulting in a lethal effect for the parasite.

145 In conclusion, the results of the present study suggest that the adulticide effect of the
146 association of IVM and DOXY is not due to a higher drug concentration of DOXY in the
147 combination protocol, nor to a lack of efficacy in the removal of *Wolbachia* from the worm
148 tissue. Future studies should concentrate on the parasite target, perhaps through in vitro
149 treatment of microfilariae with one or both drugs in order to evaluate drug concentration
150 and expression of cell detoxification mechanisms.

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200 **Table 1**

201 Treatment protocols in *D. immitis*-experimentally infected dogs (Bazzocchi et al., 2008).

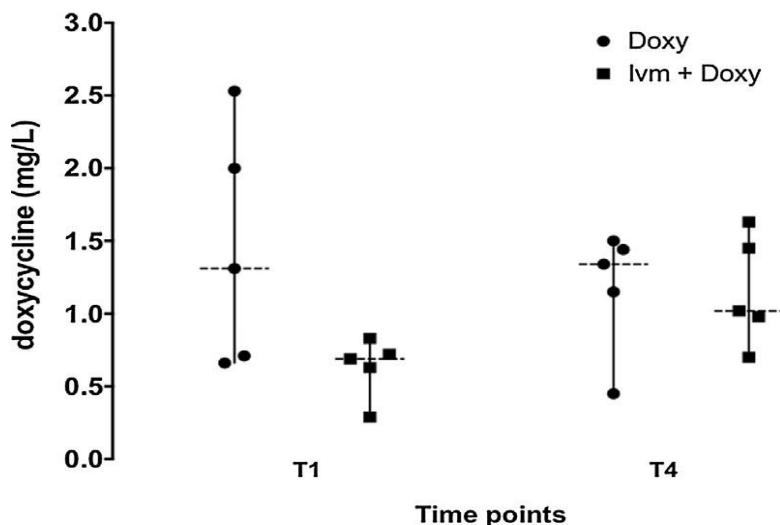
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Group	Treatment (weeks post-infection)	
	Doxycycline (10 mg/kg)	Ivermectin (6 g/kg)
DOXY	Weeks 0–6, 10–12, 16–18, 22–26, 28–34	—
IVM + DOXY	Weeks 0–6, 10–12, 16–18, 22–26, 28–34	—
CONTROL	—	—

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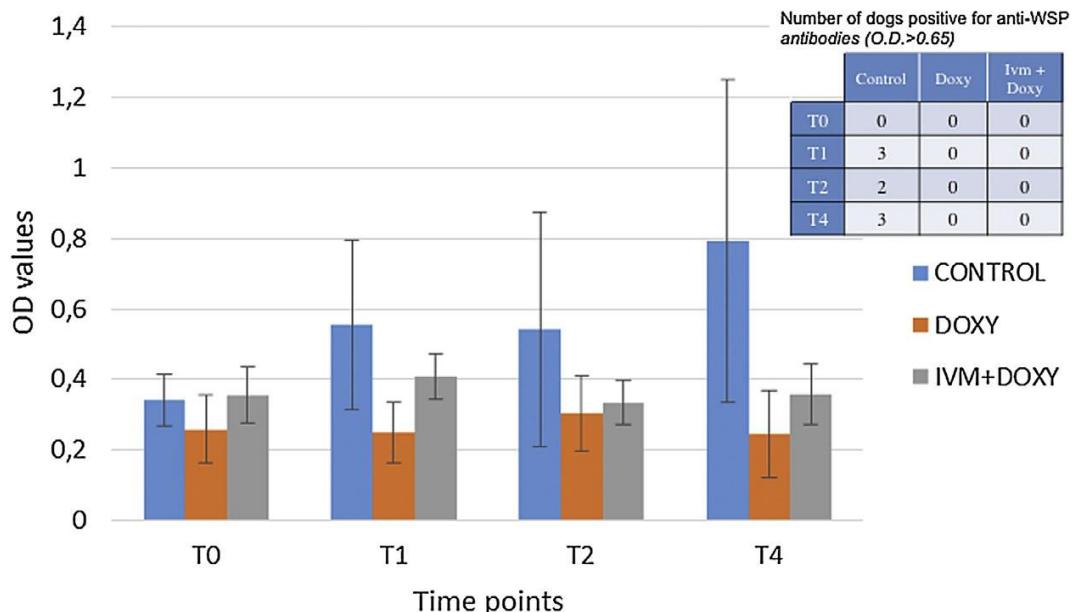


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209 **Fig. 1.** Serum concentrations of DOXY (mg/L) in HW-infected dogs (5 per group) treated
 210 with DOXY alone or with the combination IVM + DOXY, measured at time points T1 (6
 211 weeks p.i.) and T4 (34 weeks p.i). The graph shows the distribution of individual DOXY
 212 levels around the median value for the two time points.

213



214

215

216 **Fig. 2.** ELISA results for rWSP in HW+ dog sera. T0: 6 weeks pre-infection; T1 (6 weeks
 217 post infection; p.i.); T2 (10 weeks p.i.); and T4 (34 weeks p.i.). Control: O.D. values
 218 obtained from the five dogs of control group (not treated); DOXY: O.D. values obtained
 219 from the five dogs of the group treated with DOXY alone; IVM + DOXY: O.D. values
 220 obtained from the five dogs of the group treated with the drug combination. Bars indicate
 221 the means \pm SD. Cut-off value: 0.65. Table insert reports the number of WSP* dogs from
 222 each group at the different time points.