

Heart Rate Variability in Dairy Cattle: Measurement and Time Domain Analysis

M. Minero, E. Canali, V. Ferrante, C. Carenzi.

M. Minero, DVM, **E. Canali**, DVM, **V. Ferrante**, DVM, **C. Carenzi**, DVM, Istituto di Zootecnica Veterinaria, Università degli Studi di Milano, Via Celoria, 10, 20133, Milano, Italy.

1 The individual differences and the physiological state of animals
2 determine their vulnerability to stress and cause different subjects to react
3 in several ways to the same stressors (Porges, 1985). So, it is of particular
4 interest, in intensive farm conditions, to identify those physiological
5 characteristics which predispose animals to succeed or not in coping with a
6 potentially stressful environment (Terlow et al, 1997).

7 It is known that central mechanisms take part in the modulation of
8 challenge reactions and stress sensibility: both branches of the Autonomic
9 Nervous System respond rapidly to central influences, but only an
10 enhancement in sympathetic tone and changes in endocrine parameters
11 have generally been associated with the stress response in farm animals
12 (Broom and Johnson, 1993). Relatively little interest has been directed to
13 the Parasympathetic role in defining stress, but there is some evidence that
14 in mammals the vagal component of the Autonomic Nervous system plays a
15 major role in regulating heart rate in response to stressors and could be
16 considered a physiological marker of stress vulnerability (Hansen and von
17 Borrell, 1998; Hopster and Blokhuis, 1994; Hull et al, 1990; Porges, 1995;
18 Sgoifo et al., 1997). Cardiac vagal tone may be non invasively evaluated by
19 analysing the Heart Rate Variability (HRV) which gives information about
20 the sympathetic-parasympathetic autonomic balance (van Ravenswaaij-Arts
21 et al, 1993). Porges (1995) proposed that the physiologic variability in heart
22 rate should be considered a better indicator of the individual capacity to
23 cope with the environment than simple heart rate and Hopster and Blokhuis
24 in 1994 pointed out the importance of expanding knowledge of HRV in farm
25 animals. HRV, which is the conventionally accepted term to describe
26 variations of both instantaneous heart rate and interbeat intervals (Task
27 Force of the ESC and the NASPE, 1996), has been quantified in several lines
28 of human clinical research since 1965 when Hon and Lee noted that foetal
29 distress was preceded by a reduction in HRV. Later in the 1980s this
30 parameter was confirmed to be a reliable predictor of life-threatening
31 events in patients with cardiac disorders and it has only recently been

1 investigated as a measure of physical and psychological strain (Task Force
2 of the ESC and the NASPE, 1996).

3 Most of the studies conducted on animals considered laboratory rodents,
4 dogs and miniature swine as animal models for biomedical research while
5 horses gained attention because of their predominant parasympathetic
6 nervous activity at rest (de Jong et al, 2000; Kuwahara et al, 1999). Up to
7 now studies on farm animals have been scarce probably due to the practical
8 difficulties encountered performing reliable measures of HRV in field
9 conditions. The first step in analysing HRV is actually to achieve reliable
10 recordings of consecutive heart beats and this can be obtained by Holter
11 monitoring the animals or recording every interbeat interval (Fig.1) with
12 heart rate monitors suited to this purpose (Calvert, 1998). In cattle, the
13 heart rate has been recorded during different procedures (Lay et al, 1992;
14 Le Neindre, 1989) and Polar^R heart rate monitors were successfully
15 validated for use with dairy cows (Hopster and Blokhuis, 1994). However in
16 field conditions the presence of artifacts caused by movements of electrodes
17 on the skin or muscle contraction could often impair the obtaining of beat to
18 beat intervals suitable for HRV.

19 The variation in heart rate may then be evaluated in two ways: calculating
20 mathematical indices in the time domain analysis or studying frequency
21 specific oscillations of R-R intervals (in an ECG record, the duration of time
22 between adjacent QRS complexes, Fig.1) in frequency domain analysis. The
23 time domain variables give quantitative information on how much variability
24 there is, moreover, various R-R interval distribution histograms and
25 Poincaré graphs give fairly accurate qualitative information on HRV at a
26 glance (Calvert, 1998).

27 The aims of this research were: to develop standards of measurement for
28 farm evaluation of HRV in dairy cattle and to assess HRV in a group of
29 animals of the same gender and different age.

30 Heart Rate Variability (HRV) of 6 female calves, 6 heifers, 6
31 primiparous and 6 cows homogenous in age, with no clinical signs of cardiac
32 diseases, was monitored.

1 The animals were group stabled, according to age, in a deep litter system,
2 and belonged to an intensive dairy farm near the city of Milan (Italy). The
3 cows were fed unifeed *ad libitum*. Water was available *ad libitum*. HRV was
4 measured using a non-invasive telemetric system (Polar Vantage NV)
5 settled to record every interbeat interval. The system consisted in two
6 special electrodes adapted for large animals (Horse Trainer) connected with
7 a wire to a transmitter. The heart rate signal was transmitted to a wrist
8 receiver and all the apparatus was attached to a girth belt. Positioning the
9 girth belt just behind the scapula of the bovines, one of the two electrodes
10 was placed on the cardiac area and the other one on the upper left thorax,
11 just behind the scapula. All the animals were always handled by the same
12 person and they were first well accustomed to the belt. When each
13 recording session was completed, the belt was removed and the heart rate
14 data were uploaded into a notebook for the analysis. HRV was recorded
15 during different days, in the morning, each session lasting 15 minutes while
16 animals were restrained in their usual pens, in self-tethering gates. For each
17 animal, the indices were calculated in a time window of five minutes,
18 chosen from the whole period of recording, because data to be compared
19 should have the same duration, and five minutes seem to be an appropriate
20 option for the time domain methods (van Ravenswaaij-Arts et al, 1993;
21 Task Force of the ESC and the NASPE, 1996). HRV was assessed by
22 calculation of indices based on statistical operations on R-R intervals (time
23 domain analysis). The following heart rate variability indices measured in
24 the time domain area and Poincaré graphs were calculated using the Polar^R
25 software (Tab.1): **SD**, standard deviation of the R-R intervals; **MAXMIN**,
26 difference between maximum and minimum R-R interval; **RMSSD**, root
27 mean square of successive differences; **Poincaré graph**, in this graph
28 successive R-R intervals are plotted against each other; **INDA**, in the
29 Poincaré graph index A is the diameter that is parallel with the diameter of
30 the co-ordinate system. It represents the standard deviation of continuous
31 long-term RR-interval variability; **INDB**, index B is transverse to index A,

1 represents the standard deviation of instantaneous beat-to-beat RR-
2 interval; **INDA/INDB**, indexA:indexB ratio.

3 HRV indices in different classes of age were analysed by analysis of variance
4 ANOVA (SAS, 1985).

5 Abnormal waveforms in R-R intervals were found in the recordings
6 referring to technical artifacts. Working with animals in field conditions, the
7 poor contact between electrodes and skin and the movement of electrodes
8 due to the presence of hair and of cutaneous muscles could have caused the
9 presence of the artifacts as also reported in literature (Hopster and
10 Blokhuis, 1994; de Vel, 1984). Nevertheless the equipment and the
11 standards of measurement used obtained acceptable recordings of interbeat
12 intervals with less than 5% of artifacts in every analysed interval, meeting
13 the requirements of the ESC and the NASPE for the human clinical
14 evaluation of HRV (Task Force of the ESC and the NASPE, 1996). To analyse
15 HRV over longer periods of time (24 hrs), efforts should be made to further
16 improve the accuracy of recording equipment and editing processes of the
17 data because the presence of even few artifacts in the analysing data may
18 lead to serious misinterpretations of results. In other research carried out in
19 more controlled conditions, more sophisticated equipment which recorded
20 data more accurately was used (de Jong et al, 2000; Kuwahara et al,
21 1999). However, the use of a non-invasive telemetric system is essential in
22 farm conditions to give the researchers insight into Autonomic Nervous
23 System control not accessible with restrictive, invasive procedures greatly
24 influencing animal behaviour in that situation.

25 The Poincaré plot of Fig.2 qualitatively illustrates the HRV of one of the
26 cows. Sudden changes in heart beat are shown as points far from the
27 others. Low levels of HRV are represented by a narrow ellipse drawn around
28 the area covered by the points while a higher HRV would give larger
29 ellipses.

30 Table 1 shows the values of the time domain indices observed in the
31 animals grouped by age. The level of HRV presented a great individual
32 variability, evidenced by high standard deviations, suggesting that in further

1 studies it would be best to perform intra-individual comparisons of indices
2 during different challenges and breeding conditions. The analysis of time
3 domain indices of the heart period data indicated that the HRV of the
4 observed animals decreased, with no statistical difference (INDA and
5 MAXMIN $P=0.09$), with advancing age. This pattern is consistent with the
6 studies conducted in humans where the amount of HRV is influenced by
7 maturational factors. In adults an attenuation of respiratory sinus
8 arrhythmia can be observed while the decline of HRV starts in childhood
9 (van Ravenswaaij-Arts et al, 1993).

10 Considering the effect of age on the HRV of the animals in this study, larger
11 population studies with longitudinal follow-ups are needed to establish
12 normal standards of HRV for different ages, sexes and environment. As
13 already pointed out in human clinical research, this uncertain knowledge
14 limits the interpretation possibilities of HRV, especially when comparing data
15 from different subjects and could lead to incorrect explanation of the results
16 (Task Force of the ESC and the NASPE, 1996). The studies carried out up to
17 now in order to evaluate the possibility of using HRV as a stress indicator in
18 animals have shown controversial results and this was probably due to the
19 use of animals of different ages (Forde and Marchant, 1999; de Jong et al,
20 2000). The results of this research indicate that HRV is an interesting and
21 potentially reliable indicator for the assessment of welfare at farm level.
22 Further studies are needed and should consider both intra-individual
23 differences in HRV of animals exposed to various procedures and large
24 population basal recordings to better determine the predictive value of HRV
25 in the identification of individual vulnerability to stress.

ACKNOWLEDGMENTS

We are grateful to Dr. M. Bruni for giving access to his animals and facilities and to Dr. A. Loaldi for helping in collecting data.

REFERENCES

BROOM, D.M. & JOHNSON, K.G. (1993). Stress and Animal Welfare. London, Chapman & Hall pp. 92-107.

CALVERT, C.A. (1998). Heart Rate Variability. *Veterinary Clinics of North America: Small Animals Practice* **28** (6), 1409-1427.

FORDE, R.M. & Marchant, J.N. (1999). Heart Rate Variability: a novel non-invasive means of assessing fear responses in animals ?. Proceedings of the 33rd congress of the ISAE, Lillehammer, Norway.

HANSEN, S. & VON BORRELL, E. (1998). Impact of pig grouping on sympatho-vagal balance as measured by heart rate variability. Proceedings of the 32nd congress of the ISAE, Clermont-Ferrand, France.

HON, E.H. & LEE, S.T. (1965). Electronic evaluations of the foetal heart rate patterns preceding foetal death: further observations. *American Journal of Obstetrics and Gynaecology* **87**, 814-826.

HOPSTER, H. & BLOKHUIS, H.J. (1994). Validation of a heart-rate monitor for measuring a stress response in dairy cows. *Canadian Journal of Animal Science* **74**, 465-474.

HULL, S.S., EVANS, A.R., VANOLI, E., ADAMSON, P.B., STRAMBA-BADIALE, D.E., ALBERT, D.E., FOREMAN, R.D. AND SCHWARTZ, P. J. (1990). Heart rate variability before and after myocardial infarction in conscious dogs at high and low risk of sudden death. *Journal of American College of Cardiology* **16**, 678-685.

DE JONG, I.C., SGOIFO, A., LAMBOOIJ, E., KORTE, S.M., BLOCKHUIS, H.J. AND KOOLHAAS, J.M. (2000). Effects of social stress on heart rate variability in growing pigs. *Canadian Journal of Animal Science* **80**, 273-280.

KUWAHARA, M., HIRAGA, A., KAI, M., TSUBONE, H. AND SUGANO, S. (1999). Influence of training on autonomic nervous function in horses: evaluation by power spectral analysis of heart rate variability. *Equine Exercise Physiology* **30**, Suppl., 178-180.

LAY, D.C., JR, FRIEND, T.H., RANDEL, R.D., BOWERS, C.L., GRISSOM, K.K. AND JENKINS, O.C. (1992). Behavioural and physiological effects of freeze or hot-iron branding on crossbred cattle. *Journal of Animal Science* **70**, 330-336.

LE NEINDRE, P. (1989). Influence of rearing condition and breed on social behaviour and activity of cattle in novel environments. *Applied Animal Behavioural Science* **23**, 129-140.

PORGES, S.W., (1985). Spontaneous Oscillations in Heart Rate: Potential Index of Stress in Moberg G.P. (ed.) *Animal Stress*. Bethesda, American Physiological Society pp. 97-111.

PORGES, S.W., (1995). Cardiac vagal tone: A physiological index of stress. *Neurosciences and Biobehavioral Reviews* **19** (2), 225-233.

VAN RAVENSWAAIJ, C.M.A., KOLLÉE, L.A.A., HOPMAN, J.C.W., STOELINGA, G.B.A. AND VAN GEIJN, H.P., (1993). Heart Rate Variability. *Annals of Internal Medicine* **118** (6), 437-445.

SAS (1985). User's Guide. Cary, Sas Institute Inc.

SGOIFO, A., DE BOER, S.F., WESTENBROEK, C., MAES, F.W., BELDHUIS, H., SUZUKI, T. AND KOOLHAAS, J.M. (1997). Incidence of arrhythmias and heart rate variability in wild-type rats exposed to social stress. *American Journal of Physiology* **273**, H1754-H1760.

TASK FORCE OF THE EUROPEAN SOCIETY OF CARDIOLOGY AND THE NORTH AMERICAN SOCIETY OF PACING AND ELECTROPHYSIOLOGY, (1996). Heart Rate Variability: Standards of Measurement, Physiological Interpretation and Clinical Use. *Circulation* **93** (5), 1043-1065.

TERLOW, E.M.C., SCHOUTEN, W.G.P. AND LADEWIG, J, (1997). Physiology in Appleby M.C. and Hughes B.O. (eds.) *Animal Welfare*. Wallingford, CAB International pp. 143-158.

DE VEL, O.V., (1984). R-wave detection in the presence of muscle artifacts. *IEEE Transaction on Biomedical Engineering* **31**(11), 715-717.

Indices	Description	Significance	Calves	Heifers	Primiparous	Cows
SD	Stdev of RR intervals	Estimate of overall HRV	127.3±187.2	51.4±21.7	29.2±9.6	33.1±11.4
INDA	Longer diameter of the Poincaré graph	Long term HRV, sympathetic activity	141±226.9	68.8±29.9	39.8±13	44.9±16.3
INDB	Shorter diameter of the Poincaré graph	Short term HRV, parasympathetic activity	79.1±152.2	20.8±10.2	10.4±5	11±4.3
MAXMIN	Difference between max and min RR intervals	All components contributing to variability	443.5±326.6	343.5±229	208.7±78.1	246.5±126.1
RMSSD	Root mean square of successive differences	Estimate of short term components of HRV	112.2±215	29.8±14.9	14.8±7	15.7±6.2
INDA/INDB	IndexA/indexB ratio	Sympatho-vagal balance	3.8±3.6	3.6±1.6	4.2±1.3	4.6±2.1

Tab.1: Mean (stdev) HRV indices in different ages.

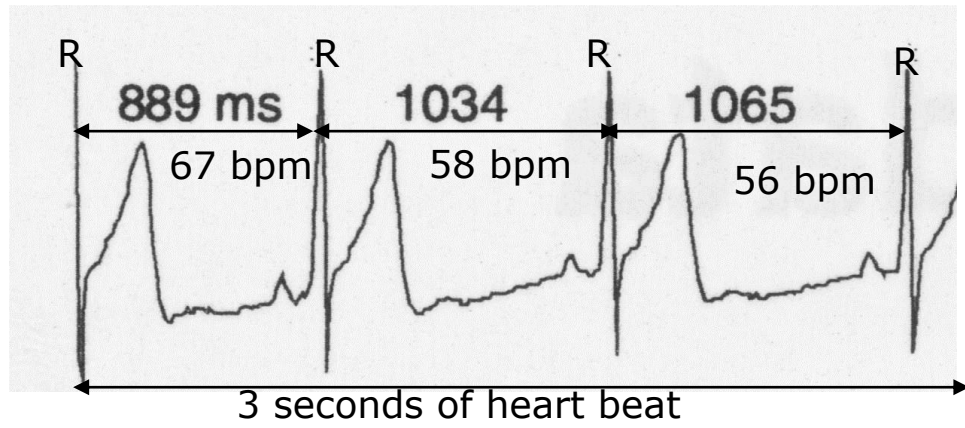


Fig.1. Measure of the R-R interval changes in heart rate

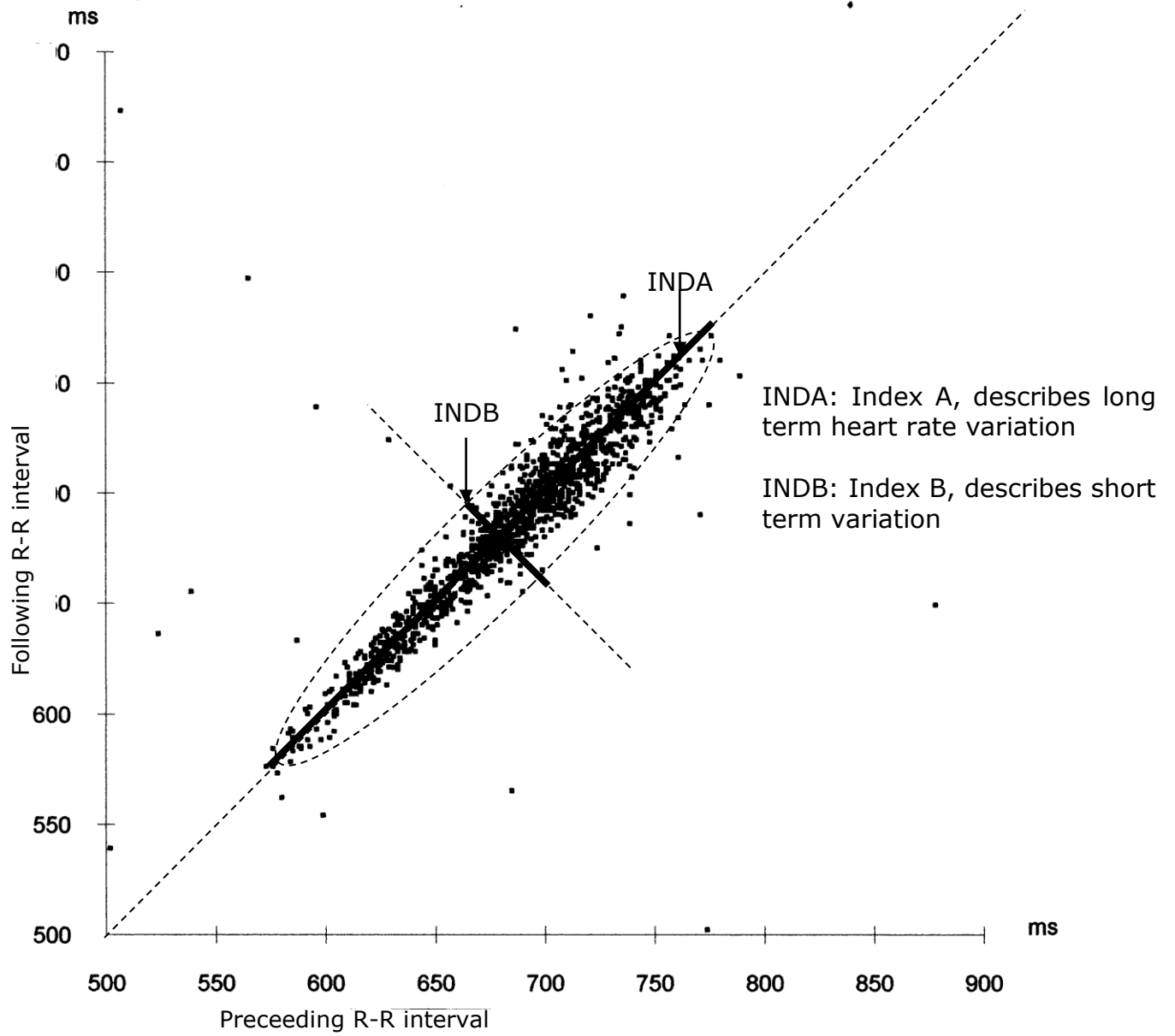


Fig.2: Poincaré plot used to illustrate heart beat rhythm and its changes in a cow.