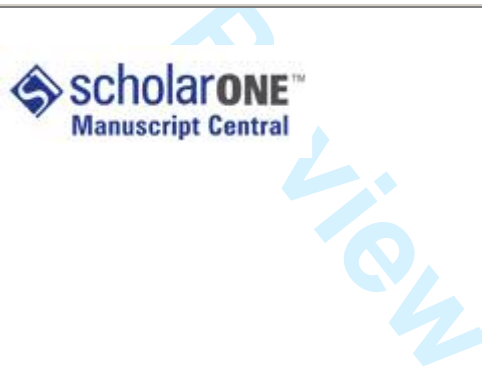


Evaluation of a bio-coating as a solution to improve barrier, friction and optical properties of plastic films

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Evaluation of a bio-coating as a solution to improve barrier, friction and optical properties of plastic films

By Stefano Farris*, Laura Introzzi and Luciano Piergiovanni

diSTAM, Department of Food Science and Microbiology, University of Milan, Via Celoria 2 - 20133 Milano, Italy

Abstract

The present research dealt with evaluating barrier, friction and optical properties of three different plastic films after deposition of a gelatin-based bio-coating. The composite films showed improved barrier properties against oxygen and UV radiation. The oxygen transmission rate decreased in the order of 73% for oriented polypropylene (OPP), 56% for low-density polyethylene (LDPE) and 40% for polyethylene terephthalate (PET). The increased UV-barrier characteristics ranged from 20% for OPP to 12% for both LDPE and PET. Static and kinetic coefficients of friction were significantly decreased both in the film-to-film and in the film-to-metal tests, leading to a desirable value for many applications. However, bio-coated films showed lower optical performances in terms of transparency and haze. Transparency decreased mainly for LDPE (36%), whereas the haze index increased especially for OPP (85%). Not significant differences were observed as far as the water vapour permeability is concerned, except for a slight reduction for PET (from $15.78 \text{ cm}^3 \text{ m}^{-2} \text{ day}^{-1}$ to $13.53 \text{ cm}^3 \text{ m}^{-2} \text{ day}^{-1}$ at 23°C and 90% of RH), suggesting that not meaningful effects arose from the addition of a hydrophobic component in the original formulation. Finally, the solubility in water of the coating was around 25% for all the three plastic substrates. The obtained data suggest that the lipid-protein coating tested in this study, in spite of its great potential for enhancing some characteristics of plastic packaging films, still exhibits negative aspects which necessitate further improvement.

KEY WORDS: Barrier properties; bio-coating; gelatin; coefficient of friction; plastic films; UV transmission.

*Correspondence to: S. Farris, Department of Food Science and Microbiology, University of Milan, Via Celoria 2 - 20133 Milano, Italy

Email: stefano.farris@unimi.it

INTRODUCTION

Over the last decade, the interest in using natural resources in order to replace, totally or partially, synthetic packaging has increased, as shown by the number of research papers, reviews books and book chapters reported in the scientific literature.¹⁻⁶ Along with the efforts made to preserve high food quality, worldwide attention focused on decreasing plastic waste to improve environmental standing.⁷ The amount of food plastic packaging material is high and one option to replace it is the use of biopolymers, for example in the form of films or coatings. Within bio-based materials, edible films and coatings have received huge attention in the recent years.⁸⁻¹⁰ In fact, films made from polysaccharides, proteins and lipids have shown properties similar to those of plastic films. In addition, they offer some potential advantages represented by the availability, low cost and biodegradability. These characteristics meet the consumers' trends', leading to a greater attention to both what is natural and environmental compatible. Furthermore, for industry, this provides innovative and economic solutions. For these reasons, biopolymers have a potential use as sustainable food packaging materials.

According to these considerations, the application of edible coatings directly on different food items has been deeply investigated by many authors with positive results.¹¹⁻²⁰ Furthermore, it is currently acknowledged by researchers that replacing food plastic packaging using a fully standing-alone edible film represents a difficult task to achieve. The major hurdle in this sense is the inability to maintain structural and functional integrity over time, especially due to the high water vapor sensitivity and poor mechanical properties of such structures.⁷ Nevertheless, a first step to use biomacromolecules as food packaging materials could be represented by a combination of both components (plastic and biopolymers), by the deposition of a thin layer on a plastic web. Currently, common plastic films are coated or laminated with synthetic polymers in the form of thin layers in order to increase specific properties of the plastic substrate. For example, polymers like poly vinyl alcohol (PVOH) or ethylene vinyl alcohol (EVOH) are used to achieve high barrier properties against oxygen, whereas polyvinylidene chloride (PVDC) is used both as an excellent water vapour

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3 and oxygen barrier. Recently, Si-derived coatings seem to offer the best performances in terms of
4 barrier and optical properties. Other meaningful techniques are plasma deposition and metallization.
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8 However, each of these surface treatments involves negative aspects, for example costs, technical
9 efforts and environmental impact. For these reasons, the use of biodegradable matrices may
10 represent a valid opportunity in the substitution of the synthetic coatings.
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15 At the present time, very few papers have been published in this field.²¹⁻²⁵ They have shown that
16 the deposition of thin bio-layers on common plastic films has a great potential. In fact, they could
17 be used for replacing the synthetic polymers utilized in the existing industrial coating processes as
18 well as to reduce the thickness of the plastic substrates attaining the same overall performances.
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20 However, in work to date only single biomacromolecular coating formulations (i.e., proteins or
21 polysaccharides) have been investigated. Moreover, authors have focused specifically on the effect
22 of the biopolymeric layers in the enhancement of the oxygen barrier property of the plastic films.
23
24 To cover the industrial requirements for new solutions, the use of a two- or multi-component
25 coating indeed could represent an alternative path. In this sense, the use of matrices with different
26 characteristics in the coating formulation could improve some performances of the final composite
27 films, for example in terms of barrier, mechanical and optical properties. Thus, to fill the existing
28 gap, this paper examined the use of a lipid-protein matrix as a thin coating on three different plastic
29 films widely used in the food packaging industry. In particular, pigskin gelatin was used as a protein
30 due to its abundance, relatively low cost, filmogenic and foam properties, high barrier properties
31 against oxygen, transparency.^{26,27} This ingredient has been previously used for food applications
32 and also for biomedical purposes to create spheres and capsules for controlled drug delivery.^{28,29} In
33 the present paper, pigskin gelatin was selected to provide the barrier against oxygen and the
34 necessary strength to the biopolymeric layer. An acetylated monoglyceride was chosen as a lipid, in
35 the attempt to provide both best water vapour barrier performances and friction properties of the
36 final composite films.³⁰ Moreover, its amphiphilic characteristic should guarantee the adequate
37 interaction with the hydrophilic component, avoiding any kind of phase separation phenomenon
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3 after deposition. Finally, glycerol was used as a plasticizer to enhance the flexibility of the final
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5 structure, preventing coating cracking.
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8 The aim of the present work was to evaluate the effect of a lipid-protein bio-coating on some
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10 properties of plastic packaging films. To this purpose, barrier (oxygen and water vapour), friction
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12 (static and kinetic coefficients) and optical (haze, transparency and UV transmission) properties of
13
14 three different plastic films (polypropylene, polyethylene terephthalate and low-density
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16 polyethylene) after the coating deposition were evaluated. The possibility to use these kind of layers
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18 as a potential and emerging packaging technology was also discussed.
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24 **MATERIALS AND METHODS**

25 **Materials**

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28 Pigskin gelatin powder type A, 133 Bloom (Weishardt International, Grauliet Cedex, France) was
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30 used as a protein component to make coatings. Acetic acid esters produced from monoglycerides
31
32 (Grindsted[®] Acetem 70-00 P) were provided by Danisco (A/S, Langebrogade, Denmark) and were
33
34 used as lipid component. They are made from edible, fully hydrogenated lard, in which 70% of the
35
36 free hydroxyl groups have been acetylated. Finally, glycerol (Giomavaro, Brugherio, Italy) as a
37
38 plasticizer was added in the coating formulation. Oriented polypropylene (OPP) of $20.0 \pm 0.5 \mu\text{m}$
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40 (Radici Film, S. Giorgio di Nogaro, Italy), polyethylene terephthalate (PET) of $12.5 \pm 0.5 \mu\text{m}$
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42 (Toray Saehan, Kyungbuk, South Korea) and low-density polyethylene (LDPE) of $48.0 \pm 0.5 \mu\text{m}$
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44 (Ticinoplast, Pogliano Milanese, Italy) were used as plastic substrates for the coating deposition. All
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46 films were provided by the suppliers as corona-discharge-treated materials.
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56 **Preparation of coated films**

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58 Gelatin powder (11.8% w/w) was mixed in distilled water with glycerol (3.9% w/w). The solution was
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60 heated to 75°C, in order to obtain a complete solubilization and denaturation of the protein. The pH

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3 was adjusted to the isoelectric point (pI) of the gelatin (≈ 8.5), using sodium hydroxide (NaOH) 1
4 M. The behaviour of gelatin chains is strongly influenced by the pH value. In particular, for pH far
5 from the pI an electrostatic repulsion exists between gelatin molecules, reducing the number of
6 junction zones connecting different chains. As the pH approaches the pI, electrostatic repulsion is
7 reduced, due to the net charge along the chain becoming close to 0. At this specific pH value the
8 affinity between gelatin and water is little or equal to zero, allowing a high reactivity between the
9 protein and other reactants (e.g. lipids).³¹

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20 The slurry was kept on a mixing heated plate for 1 h. The lipid component (5.9% w/w) was
21 added and mixed for 2 min with a homogenizer (Ika-Werke, Stanfen, Germany), at the speed of
22 24000 rpm. The solution was degassed using a vacuum pump, in order to avoid dissolved air. A
23 constant amount (1.0 cm³) of coating solution was used to coat the corona-treated side (24 x 18 cm)
24 of the different plastic films previously placed on an automatic film applicator (Ref. 1137, Sheen
25 Instruments, Kingston, UK) equipped with a steel horizontal bar to obtain a wet coating thickness of
26 10 μm . The coating process was performed at a constant speed of 150 mm/min. Coatings were
27 dried, firstly by using a constant and perpendicular flux of mild air ($25 \pm 0.3^\circ\text{C}$) at a distance of 40
28 cm from the applicator for 2 min. In a second step, composite films were stored under controlled
29 conditions ($23 \pm 2^\circ\text{C}$, $40 \pm 2.0\%$ RH) for 24 h. Both the neat plastic films and the composite ones
30 (i.e. plastic web and coating) were tested using five independent replicates in regards to thickness,
31 solubility, barrier and optical properties. Coefficients of friction were assessed using ten replicates.
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50 **Coating thickness**

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53 The thickness of the uncoated plastic films was measured with a micrometer (Dialmatic DDI030M,
54 Bowers Metrology, Bradford, UK) to the nearest 0.001 mm at 10 different random locations. For
55 the determination of the thickness of the biodegradable layers coated on the plastic films (Table 1),
56 a 10 x 10 cm sample was cut and weighed (M_I). The coating was mechanically removed by hot
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3 water (80°C) and the resulting base film weighed (M_2). The thickness of the coating was obtained
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5 using the following equation:³²
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7

$$l = \frac{M_1 - M_2}{\rho} \times 100 \quad (1)$$

8
9
10
11 where:

12
13 M_1 = unit total mass (plastic film and coating) (g dm^{-2})

14
15 M_2 = unit mass of the plastic film (g dm^{-2})

16
17 ρ = density of the aqueous solution (g cm^{-3})

18
19 l = thickness (μm)
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23 being $M_1 - M_2$ and ρ known. The thickness of the coating was also measured using an optical
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25 microscope (OM) (Micro Nikon Eclipse ME600 Laboratory Imaging, Nikon Instruments, Sesto
26
27 Fiorentino, Italy) at 100x magnification (Figure 1). In this case, films after storage were fixed on a
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29 rectangular steel holder and a sharp razor blade was then used in a specific way to cut them, in order
30
31 to permit the right observation of the cross section of the composite films. Finally, the thickness of
32
33 the layers was quantified using the software NIS-Element (Nikon Instruments, Sesto Fiorentino,
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35 Italy).
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43 **Barrier properties**

44 *Oxygen Transmission rate (OTR)*

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46 For the oxygen transmission rate measurement, an OPT-5000 (PBI Dansensor A/S, Ringsted,
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48 Denmark) equipped with a zirconia oxygen sensor was used, on the basis of the nearly-isostatic
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50 standard method as described by ASTM F 2622-08.³³ Samples were put in a paperboard support
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52 with a testing area of 42 cm^2 and then inserted in the instrument. The lower side of the testing area
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54 was continually flushed with the carrier gas (nitrogen, $70.0 \text{ cm}^3/\text{min}$ flux), whereas the upper side
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56 was exposed to the test gas flux (oxygen, $70.0 \text{ cm}^3/\text{min}$). This leads to a steady-state condition
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58 across the sample, so that the oxygen concentration will be the same at any given time. Due to the
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3 sample's permeability, the lower side (gas with high content of nitrogen) will be enriched with
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5 oxygen from the upper side (high concentration of oxygen). Measuring accurately and continuously
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7 the increase in oxygen concentration in the lower chamber will determine the permeability.
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9 Measurements were performed at a constant temperature ($23 \pm 0.5^\circ\text{C}$) and at five different relative
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11 humidity conditions (from 0% to 80% on both sides), in order to determine its influence on the
12
13 oxygen barrier property of the composite films. A time of 15 h was allowed for stabilization (i.e. the
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15 time needed to the film to reach equilibrium).
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21 22 *Water Vapour Transmission rate (WVTR)*

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24 WVTR was determined by water vapour transmission rate instrument Lyssy L-80 (PBI Dansensor
25
26 A/S, Ringsted, Denmark). The testing method as described by ASTM E 398-03 Standard Method
27
28 was used.³⁴ Film samples were double masked by manufacturer supplied aluminium foil masks with
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30 effective film test area 42 cm^2 to prevent the film being damaged during the test. Films were
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32 mounted onto instrumental cylinders, followed by analysis. Testing was performed at $23 \pm 0.5^\circ\text{C}$,
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34 with 100% RH for the wet chamber of the instrument and 10% for the dry one, yielding to a driving
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36 force of 90% RH. Each sample was oriented with the coated side facing the lowest humidity
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38 chamber.
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45 **Optical properties**

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48 Optical properties were measured using an UV-Vis spectrophotometer (Lambda 650, PerkinElmer,
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50 Waltham, USA) for measurements between 200 and 800 nm.
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55 *Transparency*

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58 Transparency was determined according to ASTM D 1746-88.³⁵ In particular, the transparency of
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60 both uncoated and coated films was measured in terms of specular transmittance, i.e. the
transmittance value obtained when the transmitted radiant flux includes only the light transmitted in

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3 the same direction as that of the incident flux in the range 540–560 nm. Then, the correspondent
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5 transparency value was obtained by the following equation:
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$$(T_s) = 100 I_s / I_0 \quad (2)$$

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9
10 where:

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12 T_s = specular transmittance at 550 nm

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14 I_s = light intensity with the specimen in the beam

15
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17 I_0 = light intensity with no specimen in the beam
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20 21 22 *Haze*

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24 Haze was measured in accordance with ASTM D 1003–00.³⁶ It is defined as the scattering of light
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26 by a specimen responsible for the reduction in contrast of objects viewed through it. In quantitative
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28 terms, it is the percent of transmitted light deviating more than an angle of 2.5° from the direction
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30 of the incident beam. For this reason, haze determination necessitates of a proper accessory able to
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32 trap also the diffuse transmitted light. To this purpose, a 150 mm integrating sphere (PerkinElmer,
33
34 Waltham, USA) was used, where the transmittance sample holder at the entrance of the sphere
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36 enables the measurement of light scattering films. In this way it is avoided any loss of light before it
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38 reaches the detector, because all the light which has passed through the sample is collected.
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45 46 *UV transmission properties*

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48 UV transmission properties were evaluated by collecting transmittance spectra of neat and coated
49
50 plastic films in the UV region 200–340 nm. The area under each transmittance curve was calculated
51
52 by an integration process. The smaller the area value, the greater will be the UV barrier properties
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54 for the specific film taken into account. UV transmission properties too were estimated using a 150
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56 mm integrating sphere.
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Friction properties

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3 In this work, both static (μ_s) and kinetic (μ_k) friction coefficients were measured. The former
4 represents the friction opposing the onset on relative motion (impending motion), whereas the latter
5 can be considered as the friction opposing the continuance of relative motion once that motion has
6 started.³⁷ In the case of solid-on-solid friction (with or without lubricants), these two types of
7 friction coefficients are conventionally defined as follows:
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$$\mu_s = F_s/P \quad (3)$$

$$\mu_k = F_k/P \quad (4)$$

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22 where:

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24 F_s = force just sufficient to prevent the relative motion between two bodies;

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27 F_k = force needed to maintain relative motion between two bodies;

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30 P = force normal to the interface between the sliding bodies.
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34 Both friction coefficients were measured using a dynamometer (mod. Z005, Zwick Roell, Ulm,
35 Germany), in accordance with the standard method ASTM D 1894-87.³⁸ The software 'TestXpert
36 V10.11 Master' was used for data analysis. Two types of analyses were carried out. In the first,
37 friction opposing the motion of each type of film (coated and uncoated) against itself was evaluated.
38 In the second, the motion of each type of film (coated and uncoated) on a metallic rigid surface (a
39 polished stainless steel 150 by 450 by 3 mm) was considered. This surface, other than acting as a
40 supporting base to guarantee a firm position between the moving crosshead and the force-measuring
41 device, served the purpose of simulating the friction between the plastic web and the metallic parts
42 of the equipments used during the manufacturing process.
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58 **Coating solubility (total soluble matter)**

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60 According to Gontard *et al.*,³⁹ film solubility in water was measured as a percentage of dry matter
of the bio-coating solubilized after 24 h, assuming that the plastic substrate does not dissolve at the

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3 same conditions. Composite films (plastic and coating) were kept in a desiccator containing calcium
4 chloride for 24 h. These films were cut by 10 x 10 cm, weighed, immersed in 250 cm³ distilled
5 water, sealed, and periodically agitated for 24 h at 20°C in an incubator (MCT 120, Angelantoni
6 Scientifica, Massa Martana, Italy). After this time, the same film pieces were taken out and dried at
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13 100°C for 24 h to determine the weight of dry matter. Solubility was then calculated as follows:
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$$17 \text{ Solubility (\%)} = [(dm_i - dm_f) / dm_i] \times 100 \quad (5)$$

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22 where:

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24 dm_i = initial dry matter of the composite films

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27 dm_f = dry matter of the composite films after 24 h immersion in water
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32 **Statistical analysis**

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34 Statgraphics Plus 4.0 software (STSC, Rockville, USA) was used for the one-way ANOVA, in
35 order to check for differences between and within groups (each plastic film, coated and uncoated).
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39 The mean values, when appropriate, were separated by LSD's multiple range test at $p \leq 0.05$.
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43 **RESULTS AND DISCUSSION**

44 **Barrier properties**

45 *Oxygen Transmission rate (OTR)*

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47 Table 1 shows the results obtained for the oxygen transmission rate for uncoated and coated plastic
48 films at 0% relative humidity. As can be seen, the addition of the biopolymeric layer led to a drastic
49 decrease of the oxygen permeability, ranging from 73% for the OPP to 40% for the PET. The
50 barrier effect of the coating is undoubtedly due to the presence of gelatin. This protein exhibits
51 oxygen permeability roughly close to that of PVDC and EVOH films at 0% relative humidity.^{40, 41}
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Moreover, its high linear structure makes it a grater oxygen-barrier than others globular proteins (corn zein, wheat gluten, soy protein or whey protein) due to a higher cohesive energy density and a lower free volume.⁴² However, in this work the extent of the improvement of the oxygen barrier property of the three plastic films due to the addition of the coating was lower if compared to the results achieved in similar studies.^{21,22,25} This is linked to the different thickness of the plastic film and of the coating. In fact, previous research used plastic films of around 50.0 μm thickness, whereas the coating as used in this work had a thickness not less than 6.0 μm , hence three times greater than the gelatin-based coating tested in the present study. In addition, the initial coating formulation has to be kept in mind. In contrast to previous works, a lipid component is also present in the gelatin-based coating designed for this research. Although the addition of hydrophobic molecules is view as necessary to improve the water vapour barrier property of films, its presence causes a decrease in the cohesive energy density of the protein network and a consequent increase in free volume, acting as a plasticizer.³⁰ The final result is a less tight lattice (i.e. increased interstitial space between protein molecules) and a reduced energy binding together the protein chains, thus allowing a permeant (oxygen) to pass faster across the sample. In Figures 2-4 the evolution of the oxygen transmission rate as a function of the relative humidity is reported, for both uncoated and coated films. It is interesting to observe as the uncoated OPP and LDPE barrier properties against oxygen were almost constant over the whole investigated RH range. Conversely, the OTR values of uncoated PET decreased when increasing RH. This behavior, though unusual, is typical for those polymers like PET and aromatic polyamides, and it seems due to decreases in both diffusivity (D) and solubility (S) of the permeant as a consequence of water molecules absorption by the film.⁴³ With regard to the coated films, there is a great dependence of the OTR values on relative humidity. In particular, values similar to those measured at anhydrous conditions were measured up to 40% RH for OPP ($640 \pm 15 \text{ cm}^3 \text{ m}^{-2} \text{ day}^{-1}$), PET ($75 \pm 5.8 \text{ cm}^3 \text{ m}^{-2} \text{ day}^{-1}$) and PE ($1400 \pm 15 \text{ cm}^3 \text{ m}^{-2} \text{ day}^{-1}$). Beyond this RH limit-value the increase was significant and followed an exponential trend, in agreement with previous works.^{21,22,25} Such behaviour is analogous to that of

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3 commercial synthetic polymers often used in coating or lamination process such as poly vinyl
4 alcohol (PVOH) or ethylene vinyl alcohol (EVOH). The high moisture-sensitivity limits their
5 application to food items with low water activity, unless these polymers are incorporated as inner
6 layers between polyolefins or other polymers acting as barriers against water vapour. The
7 pronounced hydrophilic nature of gelatin is by far the main factor leading to the different behaviour
8 of the films when altering the relative humidity of the system. It could be ascribed to the chemical
9 structure of gelatin, i.e. the amino acid composition. In fact, pigskin gelatin is composed of
10 approximately 45% hydrophilic amino acids like glycine, glutamic acid, serine.⁴⁴ Accordingly,
11 gelatin interacts easily with the surroundings, absorbing water from the environmental humid air
12 and therefore undergoing significant physical changes. In particular, the gelatin-water interaction
13 leads both to an increased mobility in the hydrophilic macromolecule chains and to the swelling of
14 the protein network, which are at the base of the gas diffusivity and gas solubility phenomena,
15 respectively.⁴⁵ In spite of its small thickness, the bio-coating laid on OPP, PET and LDPE
16 effectively reduced their OTR, even though only at low to intermediate RH. These results are of
17 great interest, due to the possibility to improve the oxygen barrier performances of the plastic films
18 using very thin biopolymeric layers. Moreover, the performance of the biodegradable films
19 themselves can be properly improved by different approaches, like the modification of the original
20 formulation, the use of different deposition process or promoting physical-chemical changes by
21 using, for example, specific cross-linking agents.

22 *Water Vapour Transmission rate (WVTR)*

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24 To accomplish the enhancement of the water vapour (WV) barrier property of the plastic films, a
25 lipid component (acetylated monoglyceride) was added to the glycerol/protein matrix with a ratio
26 lipid/protein = 0.5. The obtained results are displayed in Table 1. Only a small effect on the WVTR
27 of the PET-coated film was measured (\approx 15% decrease), whereas the presence of the bio-coating
28 did not lead to any significant changes in the WV barrier characteristics of OPP and LDPE films. It

could be explained considering that, at $T = 23^{\circ}\text{C}$, OPP and LDPE themselves are excellent water vapour barriers, and the addition of a coating with a low fat-to-protein ratio is not enough to affect their initial performances. The same coating laid down on the PET film, instead, had some positive effect, even if of limited size. This is because of the different nature of PET, less hydrophobic than OPP and LDPE and hence with a higher initial WVTR value. According to Anker *et al.*,³⁰ these results establish that composite films and coatings made by the emulsion technique (in which the lipid component is dispersed through the predominant network, gelatin in this case) exhibit rather poor barrier properties against water vapour, due to the fact that water molecules are still able to permeate through the hydrophilic phase (i.e. gelatin). One solution could be to move from a low fat-to-protein ratio to a high fat-to-protein ratio, in order to increase the hydrophobic nature of the coating. However, Anker *et al.*³⁰ also demonstrated that by increasing the lipid/protein ratio the WV barrier properties did not change. Therefore, further studies are necessary to elucidate the proper use of the lipid component in order to decrease significantly WVTR of the most important commercial films. To this purpose, changes in the experimental conditions and in the mechanism of addition of the lipid component to the protein matrix (e.g. lamination) might lead to better results.

Optical properties

The optical properties of plastic films for food packaging applications are of great importance not only as far as the quality preservation of the foods is concerned, but also from an aesthetic and hedonistic point of view. From one hand, for instance, it should be desirable to have films with high barrier performances against the UV radiation, in order to avoid damaging reactions (e.g. lipid oxidation, discolouration). On the other hand, industries are always looking towards new solutions dealing with transparent films, due to the increasing consumers demand to 'see through' the package. At the same time, by using transparent films, companies can also compete against each other on the basis of the appearance of the product. Due to that, the present research also dealt with the influence of a bio-coating on some optical characteristics of three common plastic films. In

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3 particular, this study examined whether the overall effect could be considered positive, in order to
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5 plan future improvements on the original coating formulation.
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10 *Transparency and Haze*

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12 Transparency is the property of a material that allows seeing through it. Transparency of the coated
13 films was inferior to that of the neat films. As shown in Table 1, the transparency values (%)
14 decreased 17% for OPP (90.46 – 75.04), 12% for PET (82.27 – 72.69) and 36% for LDPE (84.10 –
15 54.05). On the contrary, haze is the property of a material of scattering the incident light. It is linked
16 to the reduction in contrast of objects viewed through it. The deposition of the coating led to a
17 consistent increase in haze for all the three plastic films. In particular, the haze index increased
18 mainly for OPP (85.5%) followed by LDPE (78%) and PET (70%). The decrease in transparency of
19 the plastic films after coating deposition is undoubtedly attributable to the lipid component. In this
20 sense, it is worth noting the increased turbidity of the protein-based slurry after lipid incorporation.
21 According to Rhim *et al.*,⁴⁶ an increase in haze (and a consequent decrease in transparency) occurs
22 in edible films upon the addition of fatty acids (e.g. lauric, palmitic, stearic, oleic). Kim and
23 Ustunol⁴⁷ reported that a trained sensory panel perceived whey protein isolate films as significantly
24 less transparent after the addition of the fat component (candelilla wax, in this case). As highlighted
25 by Hernandez,⁴⁸ transparency and haze of both plastic and biodegradable films is primarily related
26 to their morphology and not to their chemical structure. The obtained results are due not only to the
27 irregularities at the surface level, but also to the morphological inhomogeneity of the coating
28 thickness across the width. In particular, the lipid component in the final formulation is in the form
29 of tiny drops entrapped in the gelatin network, especially after mechanical homogenization. These
30 particles promoted the scattering phenomenon of visible light through the film, hence causing an
31 increase in the haze values. From the experimental results it can be concluded that the coating
32 designed according to the proposed formulation is hardly suitable for those packaging applications
33 in which the transparency of the polymer is a required prerequisite to assure product visibility.
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3 Conversely, their practical application could involve all those packaging solutions not requiring
4 high transparency values. In any case, improvements may include the process optimization (e.g.
5 homogenization, amount, thickness), in order to enhance these optical properties.
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10 11 12 *UV transmission properties*

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15 The negative effect of light on foods (especially dairy and meat products) has been extensively
16 investigated by different researchers, as reviewed by Mortensen *et al.*⁴⁹ Foods exposed to light
17 (natural or artificial) may undergo a severe quality decay, often due to light-induced degradation of
18 macromolecules like proteins, lipids and also vitamins. The role of the UV light is not negligible
19 because even small amounts of this radiation can promote the formation of free radicals from lipids
20 leading to photolytic autoxidation. Moreover, UV light seems to have a detrimental effect on
21 vitamins and pigment degradation.^{50,51} The final result is the appearance of off-flavours and
22 significant changes in colour that preclude their marketing. Among the different approaches that
23 could prevent these changes (food formulation and processing, storage and light exposure
24 conditions), the choice of an ideal packaging appears fundamental. The problem could be easily
25 solved using metallized plastic films or aluminium foils acting as a total barrier; however, they do
26 not enable the consumers to see the quality of the foods through the package. For that reason, the
27 use of transparent material is increasing, even with risking degradation of foods packaged in this
28 kind of films. In recent years, besides the practice of modifying the composition of the internal
29 atmosphere of a package (i.e. reducing the amount of oxygen), the use of UV filters paved the way
30 for new interesting solutions. In light of all these considerations, in the present study the
31 transmission spectra of both composite and neat films were collected within the wavelength region
32 380-200 nm. The area under each transmittance curve was then collected, in order to check for
33 some differences in the UV transmission properties after the coating application. As reported in
34 Figure 5, the bio-coating led to an enhancement of the UV transmission properties for the OPP and
35 LDPE films. The major component involved in this 'blocking-UV' capacity is the protein, as widely
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3 explained by Schaich.⁵² In particular, this UV-absorbing behaviour has to be attributed to the
4 aromatic amino acids in the gelatin composition, like phenylalanine and tyrosine. In addition,
5 further evidence was given by comparing the spectra obtained from films coated in accordance to
6 the original formulation with those obtained from films coated in absence of the lipid component.
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8 As displayed by the Figure 6, the transmittance spectra of OPP coated in the absence of the
9 acetylated monoglyceride are downwards-shifted with respect to those obtained with the lipid
10 component in the formulation, confirming that gelatin is able to partially block the UV radiation,
11 whereas the lipid drops dispersed in the coating reduce this effect. To corroborate this hypothesis it
12 must be highlighted that the stearic acid in the monoglyceride is saturated, and hence it has no
13 double bonds which would normally absorb the UV-light. Although the UV barrier properties of the
14 three plastic films increased after coating deposition, it is important to stress that they have a
15 moderate impact from a practical point of view. This is due to the fact that the most UV-induced
16 degradation phenomena during food storage at manufacturing facilities and in retails stores arise
17 from artificial sources generating especially UV-a and UV-b radiation, ranging approximately
18 between 380 and 280 nm. The biodegradable coating increased significantly the UV barrier
19 properties of OPP and LDPE plastic films only for the spectral region below 240 nm (Figure 5), i.e.
20 the UV-c region including high-energy light (e.g. sunlight) emitted in a negligible amount from
21 halogen and fluorescent lamps used in the markets.⁴⁸ In contrast, the UV-blocking effect of the
22 coating without the lipid component started from 300 nm even if the greatest effect was
23 encountered starting from 250 nm. Also in this case, some further changes could be made to
24 improve this property, for example by selecting a different natural compound able to absorb the UV
25 radiation in the targeted range.
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58 Friction properties

59 The influence of the coating deposition on the overall mechanical properties of plastic films is
60 strictly related to the specific test performed. Accordingly, as reported in previous work,²⁴ if the

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3 tensile properties are examined (e. g. tensile strength, elongation at break, elastic modulus), the final
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5 results tend to have a great dependence on the substrate rather than the layer applied on it. Just the
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7 opposite occurs when surface properties such as the static and kinetic friction coefficients are
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9 measured. Different films (above all polyolefin) tend to adhere to them or to the production
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11 equipment during film processing, causing severe trouble in terms of slowdown in performance and
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13 productivity. In order to simulate the practical situation often occurring at industrial level, two
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15 different analyses of the same test were performed in this work: the sliding of parallel film surfaces
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17 over each other and the sliding of film surfaces over a metal substrate. Great differences may be
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19 observed after deposition of the thin coating on the plastic substrates (Figure 7). This is basically
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21 due to the specific friction characteristics of the coating, which are independent from its thickness
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23 and which could be affected by the type of substrate on which is spread on (e.g. the substrate's own
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25 smoothness) and by the coating uniformity, as shown by the results reported in Table 1. The overall
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27 positive effect arising from the deposition of the coating on plastic films is attributable to the lipid
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29 component, which acts as a true slip agent. In confirmation, stearic acid is often used in the
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31 amidation process to obtain primary amides (i.e. stearamide) as well as the oleic acid (C18:1) and
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33 the erucic acid (C22:1) are used to obtain oleamide and erucamide, the most popular slip agents
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35 used during extrusion processes of polyolefin.⁵³⁻⁵⁵ The obtained results appear interesting from a
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37 practical point of view, especially considering the potential industrial application. Using such
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39 coatings including a lipid component as a slip agent allows a layer of film to slide easily over
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41 another layer of film (e.g. on a roll), or over machine surfaces during film manufacture and packing,
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43 reducing the coefficient of friction (CoF). Therefore, the final result will be the increased line speed
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45 in the manufacturing process and the enhancement of the packaging machine operations, resulting
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47 in an increased output.
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Coating solubility (total soluble matter)

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3 Solubility in water of coatings prepared from biomacromolecules is an important property strictly
4 influencing its practical application. Some potential applications might require a very low water
5 sensitivity, in order to maintain over time the initial benefits (mainly physical properties) arising
6 from the coating application. For some other applications (e.g. medical, nutritional), these layers
7 may be intended as a controlled release packaging system, for example for delivering active
8 components (e.g. antimicrobials, antioxidants) in a controlled manner over time. In other cases a
9 total solubility of the film appears beneficial before product consumption, for example when
10 nutraceuticals have been incorporated in foods such as yoghurt.⁵⁶ Consequently, the study of the
11 coating solubility in water represents a preliminary step to understand its possible applications and,
12 eventually, to design new solutions. As reported in Table 1, the three different composite films
13 showed similar solubility values (around 25%). At least three different factors contribute to this
14 result. Firstly, the high hygroscopic nature of the protein and glycerol, able to interact with water
15 molecules that penetrate in the bio-polymer bulk promoting at the beginning its swelling and then
16 its dissolution, due to the wrecking of the hydrogen and Van der Waals forces between polymer
17 chains. Secondly, this process could be accelerated by the presence of the lipid component
18 entrapped in the protein matrix. As explained above, the lipid component acts as a plasticizer,
19 reducing the cohesive energy between protein chains and increasing the interstitial space between
20 protein molecules allowing water to penetrate faster. Finally, an opposite force limits the coating
21 dissolution, i.e. the strong bond between the biodegradable matrix and the substrate. In this case it is
22 important to point out the positive effect arising from the corona discharge treatment, a routinely
23 industrial practice able to impart good adhesion between two different polymers (plastic film and
24 bio-coating, in this case) and a better strength to the final composite structures as well. The partial
25 dissolution of the bio-layer suggests that the formulation assumed for this research could be used
26 for short shelf-life food applications or when the controlled release of active compounds (e.g.
27 antimicrobials, antioxidant substances) is required. In this sense, a further work is in progress in
28 order to relate the coating behaviour in contact to a medium different than water as well as to

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3 investigate which components of the original formulation are more susceptible to dissolve.
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5 Moreover, changes in solubility can be achieved by chemical-physical modifications of the coating
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7 (e.g. using cross-linking agents).
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10 11 12 **Conclusions**

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15 It is possible to modify the original properties of common synthetic films by means of thin bio-
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17 coatings deposition. However, using two different components such as protein and lipid in the same
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19 formulation does not seem at the moment the best way to promote the improvement of some
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21 important performances of plastic substrates. This is because the addition of a lipid component in
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23 the original formulation did not lead to any meaningful and significant improvement of the original
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25 characteristics, with the exception of the friction properties. The addition of the monoglyceride
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27 acetylated to the gelatin-based formulation indeed reduced some positive effects provided by the
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29 protein biopolymer, such as the optical properties (in terms of haze and transparency), the UV-
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31 absorbing capacities and likely the oxygen barrier property too. Although these negative effects,
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33 such composite layers may represent a potential way able to meet the increasing demand for more
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35 sustainable solutions, especially if some aspects will be improved on. For example, the possibility
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37 to apply macromolecules with opposite characteristics by a lamination process should be carefully
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39 investigated. In addition, to accomplish the industrial demand for low-cost and high-performance
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41 new solutions, the enhancement of the formulation is needed, starting from the systematic study of
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43 the influence of the most important factors (e.g. components and process conditions) on the final
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45 properties of the coatings. Subsequently, an optimization step can be carried out by the modification
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47 of the chemical and physical structure of the polymer according to the required performances.
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49 Finally, further investigations should be done concerning some potential aspects, like the possibility
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51 to incorporate in these coatings active compounds for controlled release purposes and their sealing
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3 The huge potential involved in this ongoing innovation appears obvious. Their application on
4 plastic films may allow the reduction of oil-derived polymers both replacing the already-existing
5 synthetic coatings and reducing the thickness of the plastic web on which they should be spread on.
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7 In addition, considering the low unit price of the raw materials as well as the possibility to use the
8 already existing coating equipments at industrial level, a significant reduction of the cost of the
9 manufacturing process is also expected.
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For Peer Review

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Table

Film		Thickness (μm)	OTR ^a ($\text{cm}^3\text{m}^{-2}\text{day}^{-1}$)	WVTR ^b ($\text{g m}^{-2}\text{day}^{-1}$)	Haze (%)	Transparency (%T)	Coefficient of Friction (CoF)				Solubility of the bio-layer (%)
							Film-to-film		Film-to-metal		
							μ_k	μ_s	μ_k	μ_s	
OPP	Uncoated	19.6 (± 1.41)	1715.67 (± 17.10)	1.09 (± 0.11)*	1.60 (± 0.04)	90.46 (± 0.25)	0.75 (± 0.05)	1.27 (± 0.22)	0.37 (± 0.04)	0.48 (± 0.04)*	25.00 (± 5.57)
	Coated	21.25 (± 1.5)	465.67 (± 47.35)	1.26 (± 0.04)*	10.97 (± 1.03)	75.04 (± 3.48)	0.17 (± 0.02)	0.52 (± 0.07)	0.21 (± 0.03)	0.47 (± 0.03)*	
LDPE	Uncoated	47.5 (± 1.45)	2623.67 (± 47.50)	1.12 (± 0.19)*	5.71 (± 0.39)	84.10 (± 0.08)	0.64 (± 0.06)	0.18 (± 0.03)	0.86 (± 0.07)	0.46 (± 0.08)*	25.08 (± 1.73)
	Coated	49.1 (± 1.52)	1159.67 (± 52.60)	1.38 (± 0.01)*	26.13 (± 2.40)	54.05 (± 2.54)	0.29 (± 0.04)	0.20 (± 0.04)	0.51 (± 0.08)	0.46 (± 0.06)*	
PET	Uncoated	12.3 (± 0.98)	112.33 (± 1.53)	15.78 (± 0.43)	3.14 (± 0.14)	82.27 (± 0.32)	0.42 (± 0.03)	0.54 (± 0.06)*	0.27 (± 0.02)	0.34 (± 0.02)	28.33 (± 4.16)
	Coated	13.85 (± 1.05)	67.10 (± 0.70)	13.53 (± 0.02)	10.49 (± 0.54)	72.69 (± 1.02)	0.22 (± 0.03)	0.58 (± 0.10)*	0.20 (± 0.02)	0.42 (± 0.03)	

*Denote a not statistically significant difference between treatments within group (type of film) at $p \leq 0.05$ (or 95% confidence interval). Standard deviation is reported in brackets.

^a 23 \pm 0.5°C; 0% relative humidity on both sides of the sample.

^b 23 \pm 0.5°C; 90% relative humidity difference between wet (lower) and dry (upper) chambers of the instrument.

Figures Legend

Figure 1. OM cross-section of polypropylene coated film.

Figure 2. Effect of relative humidity on oxygen transmission rate of uncoated and coated OPP films.

Each data point is shown as average of five replicates. Error bar indicates SD.

Figure 3. Effect of relative humidity on oxygen transmission rate of uncoated and coated LDPE

films. Each data point is shown as average of five replicates. Error bar indicates SD.

Figure 4. Effect of relative humidity on oxygen transmission rate of uncoated and coated PET films.

Each data point is shown as average of five replicates. Error bar indicates SD.

Figure 5. UV light transmittance of uncoated (1) and coated (2) LDPE, uncoated (3) and coated (4)

OPP, uncoated (5) and coated (6) PET.

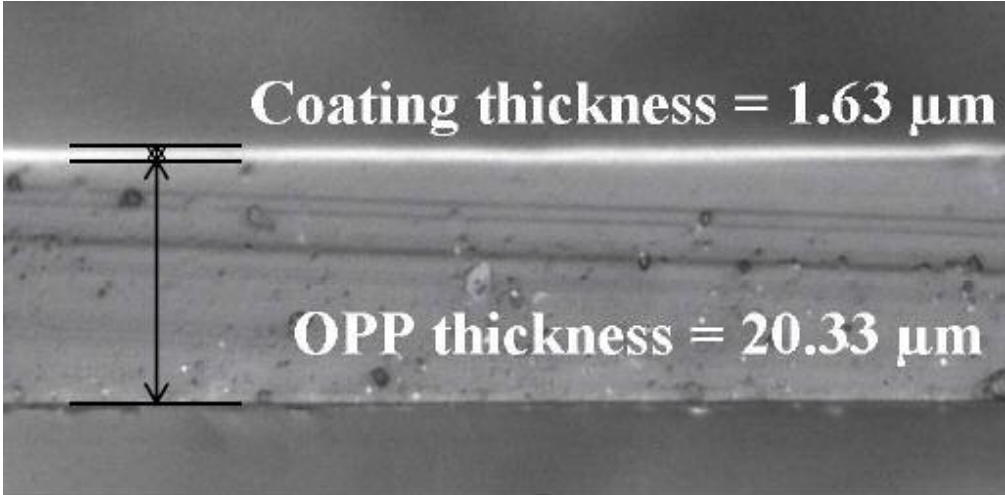
Figure 6. UV light transmittance of uncoated OPP (——), coated OPP (-----) and coated OPP

without the lipid component in the original formulation (-----).

Figure 7. Typical curves obtained for static and kinetic friction coefficients determination during the

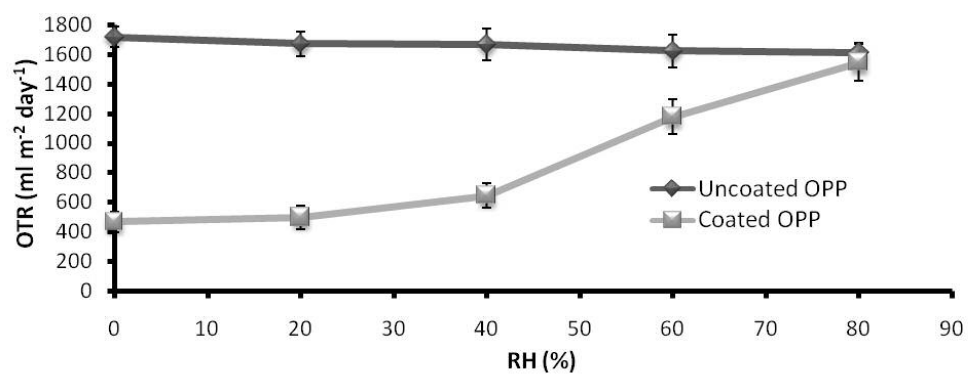
film-to-film test of uncoated and coated OPP.

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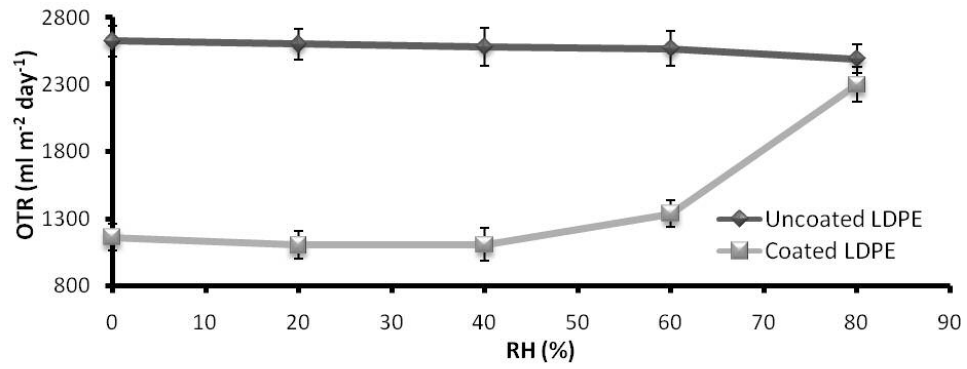
OM cross-section of polypropylene coated film.
135x99mm (96 x 96 DPI)

Review

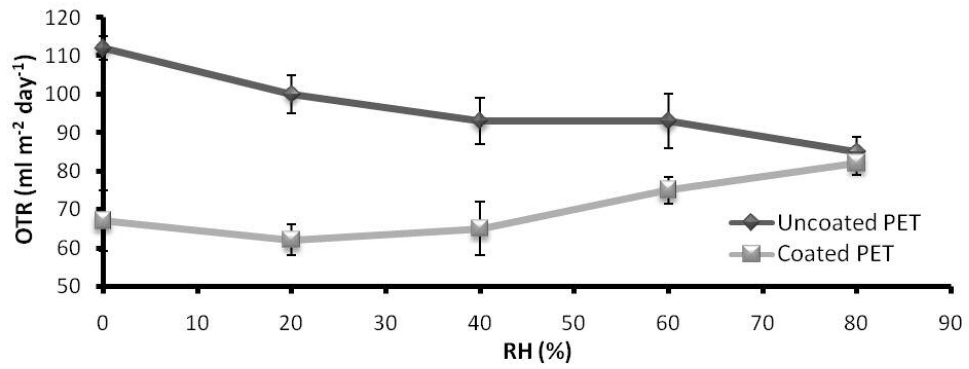


Effect of relative humidity on oxygen transmission rate of uncoated and coated OPP films. Each data point is shown as average of five replicates. Error bar indicates SD.

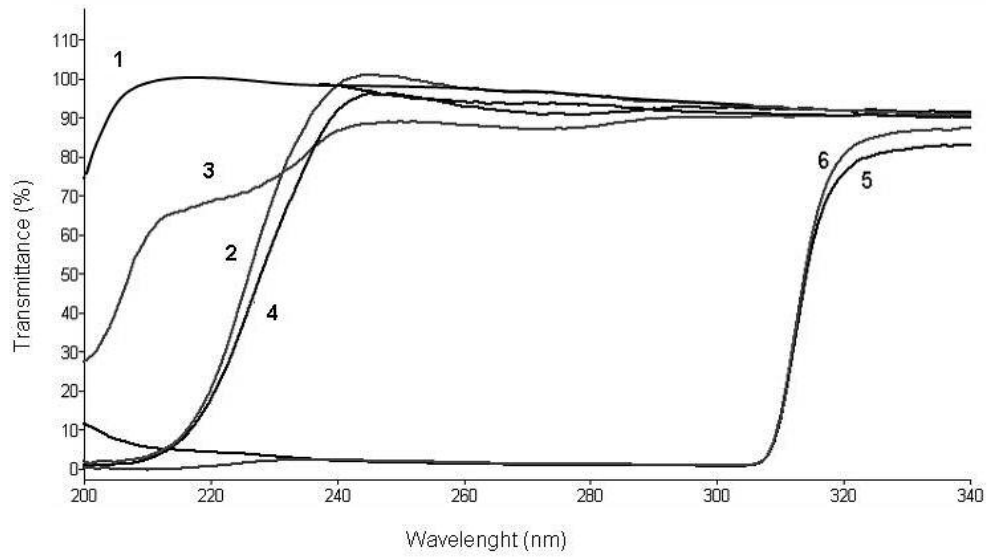
257x100mm (96 x 96 DPI)



Effect of relative humidity on oxygen transmission rate of uncoated and coated LDPE films. Each data point is shown as average of five replicates. Error bar indicates SD.
257x102mm (96 x 96 DPI)

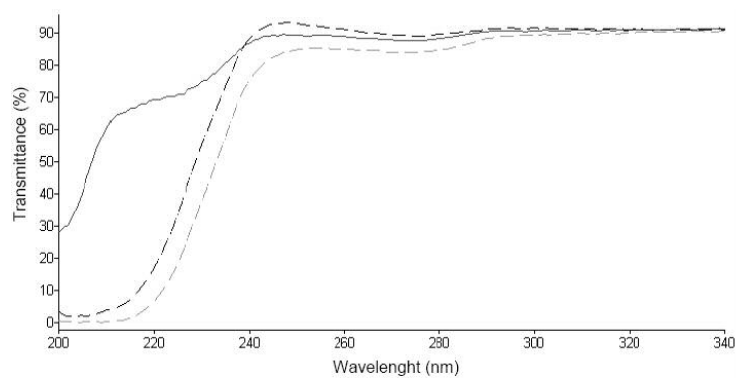


Effect of relative humidity on oxygen transmission rate of uncoated and coated PET films.
Each data point is shown as average of five replicates. Error bar indicates SD.
257x104mm (96 x 96 DPI)

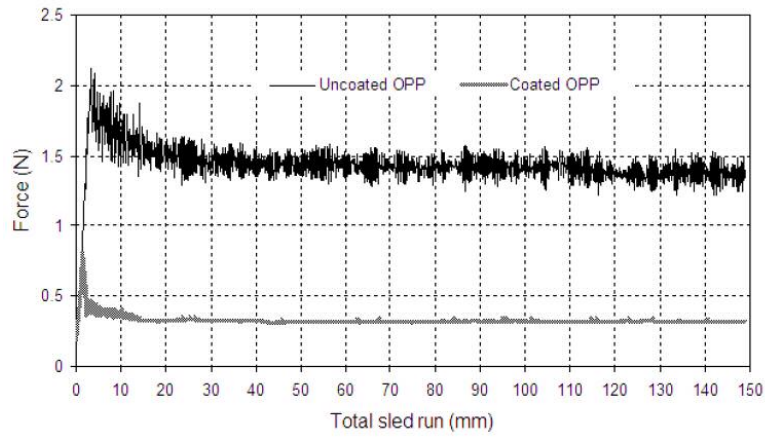


UV light transmittance of uncoated (1) and coated (2) LDPE, uncoated (3) and coated (4) OPP, uncoated (5) and coated (6) PET.
199x135mm (96 x 96 DPI)

Review



UV light transmittance of uncoated OPP (dashed line), coated OPP (dotted black line) and coated OPP without the lipid component in the original formulation (dotted grey line).
257x104mm (96 x 96 DPI)



Typical curves obtained for static and kinetic friction coefficients determination during the film-to-film test of uncoated and coated OPP.
257x115mm (96 x 96 DPI)