#### Check for updates

### OPEN ACCESS

EDITED BY Raffaella Massafra, Bari John Paul II Cancer Institute, National Cancer Institute Foundation, Scientific Institute for Research, Hospitalization and Healthcare (IRCCS), Italy

### REVIEWED BY

Yimin Cui, Peking University, China Claire Josse, University Hospital of Liège, Belgium

\*CORRESPONDENCE Sara Pizzamiglio Sara.pizzamiglio@istitutotumori.mi.it

<sup>†</sup>These authors have contributed equally to this work and share first authorship

SPECIALTY SECTION This article was submitted to Breast Cancer, a section of the journal Frontiers in Oncology

RECEIVED 26 August 2022 ACCEPTED 30 December 2022 PUBLISHED 31 January 2023

#### CITATION

Di Cosimo S, Ciniselli CM, Pizzamiglio S, Cappelletti V, Silvestri M, El-Abed S, Izquierdo M, Bajji M, Nuciforo P, Huober J, Cameron D, Chia S, Gomez HL, Iorio MV, Vingiani A, Pruneri G and Verderio P (2023) End-of-neoadjuvant treatment circulating microRNAs and HER2-positive breast cancer patient prognosis: An exploratory analysis from NeoALTTO. *Front. Oncol.* 12:1028825. doi: 10.3389/fonc.2022.1028825

#### COPYRIGHT

© 2023 Di Cosimo, Ciniselli, Pizzamiglio, Cappelletti, Silvestri, El-Abed, Izquierdo, Bajji, Nuciforo, Huober, Cameron, Chia, Gomez, Iorio, Vingiani, Pruneri and Verderio. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# End-of-neoadjuvant treatment circulating microRNAs and HER2positive breast cancer patient prognosis: An exploratory analysis from NeoALTTO

Serena Di Cosimo<sup>1+</sup>, Chiara M. Ciniselli<sup>2+</sup>, Sara Pizzamiglio<sup>2++</sup>, Vera Cappelletti<sup>1</sup>, Marco Silvestri<sup>1</sup>, Sarra El-Abed<sup>3</sup>, Miguel Izquierdo<sup>4</sup>, Mohammed Bajji<sup>5</sup>, Paolo Nuciforo<sup>6</sup>, Jens Huober<sup>7,8</sup>, David Cameron<sup>9</sup>, Stephen Chia<sup>10</sup>, Henry L. Gomez<sup>11,12</sup>, Marilena V. Iorio<sup>13</sup>, Andrea Vingiani<sup>1</sup>, Giancarlo Pruneri<sup>1</sup> and Paolo Verderio<sup>2</sup>

<sup>1</sup>Department of Advanced Diagnostics, Fondazione IRCCS Istituto Nazionale dei Tumori, Milan, Italy, <sup>2</sup>Unit of Bioinformatics and Biostatistics, Fondazione IRCCS Istituto Nazionale dei Tumori, Milan, Italy, <sup>3</sup>Breast International Group, Brussels, Belgium, <sup>4</sup>Novartis Pharmaceutical, Basel, Switzerland, <sup>5</sup>Institut Jules Bordet and l'Université Libre de Bruxelles (U.LB), Bruxelles, Belgium, <sup>6</sup>Molecular Oncology Group, Vall d'Hebron Institute of Oncology (VHIO), Barcelona, Spain, <sup>7</sup>Breast Center, University of Ulm, Ulm, Germany, <sup>8</sup>Breast Center, Cantonal Hospital St.Gallen, St. Gallen, Switzerland, <sup>9</sup>University of Leeds, Leeds, United Kingdom, <sup>10</sup>University of British Columbia, Vancouver, BC, Canada, <sup>11</sup>Department of Medical Oncology, Instituto Nacional de Enfermedades Neoplásicas, Lima, Peru, <sup>12</sup>Department of Medical Oncology, Universidad Ricardo Palma, Lima, Peru, <sup>13</sup>Department of Experimental Oncology, Fondazione IRCCS Istituto Nazionale dei Tumori, Milan, Italy

**Background:** The absence of breast cancer cells in surgical specimens, *i.e.*, pathological complete response (pCR), is widely recognized as a favorable prognostic factor after neoadjuvant therapy. In contrast, the presence of disease at surgery characterizes a prognostically heterogeneous group of patients. Here, we challenged circulating microRNAs (miRNAs) at the end of neoadjuvant therapy as potential prognostic biomarkers in the NeoALTTO study.

**Methods:** Patients treated within the trastuzumab arm (*i.e.*, pre-operative weekly trastuzumab for 6 weeks followed by the addition of weekly paclitaxel for 12 weeks; post-operative FEC for 3 cycles followed by trastuzumab up to complete 1 year of treatment) were randomized into a training (n= 54) and testing (n= 72) set. RT-PCR-based high-throughput miRNA profile was performed on plasma samples collected at the end of neoadjuvant treatment of both sets. After normalization, circulating miRNAs associated with event free survival (EFS) were identified by univariate and multivariate Cox regression model.

**Results:** Starting from 23 circulating miRNAs associated with EFS in the training set, we generated a 3-circulating miRNA prognostic signature consisting of miR-185-5p, miR-146a-5p, miR-22-3p, which was confirmed in the testing set. The 3-circulating miRNA signature showed a C-statistic of 0.62 (95% confidence interval [95%CI] 0.53-0.71) in the entire study cohort. By resorting to a multivariate Cox regression model we found a statistical significant interaction between the expression values of

miR-194-5p and pCR status (p.interaction =0.005) with an estimate Hazard Ratio (HR) of 1.83 (95%CI 1.14- 2.95) in patients with pCR, and 0.87 (95%CI 0.69-1.10) in those without pCR. Notably, the model including this interaction along with the abovementioned 3-circulating miRNA signature provided the highest discriminatory capability with a C-statistic of 0.67 (95%CI 0.58-0.76).

**Conclusions:** Circulating miRNAs are informative to identify patients with different prognosis among those with heterogeneous response after trastuzumab-based neoadjuvant treatment, and may be an exploitable tool to select candidates for salvage adjuvant therapy.

KEYWORDS

circulating microRNA, HER2-positive breast cancer, prognosis, MiR-194-5p, neoadjuvant treatment

# Introduction

Neoadjuvant therapy is progressively replacing adjuvant therapy and is emerging as a new standard of care for early-stage HER2positive and triple negative breast cancer (1). Primarily used to downstage locally advanced tumors (2), neoadjuvant therapy has the potential to enable breast cancer operability as well as to increase conservative surgery and to reduce the extent of axillary node dissection (3). Furthermore, individual response to neoadjuvant therapy provides prognostic information and assists treatment decisions after surgery in patients with human epidermal growth factor receptor 2 (HER2)-positive and triple-negative breast cancer. Indeed, while patients with a pathological complete response (pCR), defined by the absence of invasive tumor in excised breast tissue and nodes, have a favorable prognosis, those with persistent disease within the surgical specimens require adjuvant therapy escalation (4, 5).

The efficacy of additional treatments in HER2-positive breast cancer patients not achieving a pCR after neoadjuvant therapy is being investigated in several studies (6), and has been reported as beneficial by the KATHERINE trial (7). Noteworthy, not all patients with an incomplete tumor response to neoadjuvant therapy relapse two third of them are indeed disease-free at five years from surgery (8) - while up to 20% of patients who achieve a pCR eventually recur (9). This creates uncertainty, and calls into question the prognostic relevance of pCR. However, a reliable tool to separate patients at risk of relapse from those already cured after neoadjuvant therapy is currently lacking, hindering appropriate selection of adjuvant therapy escalation candidates. Primary tumor gene expression, proteomics and mutational profiling represent promising biomarkers, but need repeated tissue sampling and high-profile technology which limit their use in daily practice (10, 11). A possible alternative is offered by the development of a non-invasive procedure such as liquid biopsy. In particular, circulating microRNAs (miRNAs) are promising biomarkers due to their storage stability, easy handling, and promising expression signatures associated with treatment response (12). Baseline levels of circulating miRNA-21, -4734 and -150-5p (13, 14), or "on treatment" levels of circulating miRNA-140-5p (15) have already been associated with treatment response to HER2-targeted therapies. As it is common practice to offer post-operative treatment to non-responding patients, and pCR is not always reliable, we aimed to challenge circulating miRNAs detected at the end of neoadjuvant therapy as prognostic biomarkers. To this end, and given that most patients with HER2-positive early-stage breast cancer, in the face of different strategies to increase or decrease systemic therapy, continue to receive only chemotherapy and trastuzumab, we decided to analyze the association between pre-operative circulating miRNAs and eventfree survival (EFS) of patients treated in the NeoALTTO trial (16) with trastuzumab-based therapy.

# Materials and methods

### Patients

This is an exploratory analysis of the multicenter phase III NeoALTTO study (16), which randomized patients with HER2positive primary breast cancer >2 cm to lapatinib (n = 154), trastuzumab (n=149), or their combination (n=152) for 6 weeks, followed by paclitaxel for 12 weeks. Surgery was performed within four weeks from the last paclitaxel dose. After surgery, patients received fluorouracil, epirubicin, and cyclophosphamide for 3 cycles and continued the same anti-HER2-targeted agent of the neoadjuvant phase to complete 52 weeks of treatment. NeoALTTO primary endpoint was pCR; secondary endpoints included EFS, defined as the time from randomization to first event. As reported (15, 16), all enrolled patients signed the main study consent form, which included a non-specific clause for using blood samples collected at baseline, during treatment, prior to surgery, and eventually at the time of relapse for future research. The current analysis was approved (INT 186-13) by the Ethics Committee of Fondazione IRCCS Istituto Nazionale dei Tumori, Milano.

## Sample collection

Patients randomized to trastuzumab arm and with an available plasma sample collected prior to surgery were considered suitable for the purpose of this study, designed to address the prognostic value of end-of-neoadjuvant treatment circulating miRNAs. A training-testing approach was used for model building and confirmation, respectively.

### Circulating miRNA profiling data processing

Blood samples, collected in BDTM P-100 tubes (BD Bioscience), were separated within 2 hours of collection into plasma aliquots by centrifugation (2000-3000g for 15 minutes at room temperature) and stored at -80°C until assayed at the central biobank of Vall d'Hebron University Hospital (Barcelona, Spain). Plasma samples were shipped to Fondazione IRCCS Istituto Nazionale dei Tumori for RNA isolation as already reported (15). Briefly, reverse transcription and circulating miRNA profile was performed using the miRCURY LNA<sup>TM</sup>Universal RT microRNA PCR system according to the Exiqon manufacturer's instructions. A total of 752 circulating miRNAs were profiled using microRNA Ready-to-Use PCR, Human panel I+II in each sample. The amplification curves were analyzed using the Roche LC software for determination of quantification cycle (Cq) values. Consistently with our previous report (15), we considered background filtered (BF) Cq data as processed by Exiqon (i.e., for assays that do not yield any signal on the negative control, the upper limit of detection was set to Cq = 37; otherwise, it was set to 3 Cq lower than the Cq value of a negative control, 17). The BF Cq values were then processed to calculate the relative quantity (RQ) of each miRNA by using the comparative threshold cycle method (18) following the formula  $2^{-\Delta Cq}$  (where  $\Delta Cq=Cq_{miRNA}$  - Cq<sub>reference</sub>). The Cq<sub>reference</sub> was computed according to the overall mean approach (19).

### Statistical analysis

In the training set the association between circulating miRNAs levels (RQ considered on log2 scale) and event free survival (EFS) was assessed by resorting to a univariate Cox regression model. Event-free survival was defined as time from randomization to first event (i.e., events were defined as breast cancer relapse after surgery, second primary malignancy, death or failure to complete neoadjuvant therapy because of disease progression) (8). The relationship between each miRNA and clinical outcome was investigated by restricted cubic splines (20). In this selection step, according to the number of event per variable (EPV) (21), we considered as potentially relevant only those miRNAs detected in at least 10 patients experiencing the event of interest and in at least 85% of both training and testing sets (i.e. selected circulating miRNAs). Statistical significant circulating miRNAs at univariate analysis were included in multivariate Cox regression models following the all-subset approach (22) using penalized maximum likelihood estimations according to Firth method (23). For each model, the C-statistic (and its 95% Confidence Interval [CI]) computed according to Uno et al. (24) was used as pivotal measure for performance evaluation. Models with a statistically significant performance (i.e., with the lower 95%CI of the C-statistic >0.50) in the training set were then evaluated in the testing set. Those retaining a statistically significant performance in the testing set and including miRNAs with the same Hazard Ratio [HR] direction in both training and testing set were defined as promising prognostic signature(s). Noteworthy, models including miRNAs indicated in literature as haemolysis related (i.e., miR-16, miR-92a, miR-451 and miR-486) (25) were excluded as well as those with redundant circulating miRNAs according to the 95%CI of the Spearman correlation coefficient (i.e. upper limit of the 95%CI > 0.80 in absolute value). The "best" prognostic signature was eventually identified as that showing the highest prognostic performance in both the training and testing sets (Supplementary Figure S1). Next, by considering the whole study cohort (i.e., training and testing sets together), the prognostic performance of the "best" prognostic signature was evaluated with respect to clinicopathological variables, i.e., estrogen receptor (ER) status (negative versus [vs] positive), nodal status ( $\geq$ N1 vs N0), tumor size (>5cm vs  $\leq$ 5cm), age (≥50 vs <50) and pCR (yes vs no). A 30-week landmark analysis was performed when pCR was considered as a covariate. Finally, a Cox regression model including each selected circulating miRNA the pCR status (main effects) together with their first order interaction term was implemented to highlight circulating miRNAs differently associated with EFS depending on pCR status. All statistical analyses were carried out with the SAS (version 9.4.; SAS Institute, Inc., Cary, NC) and R software (version 3.6.0; R Foundation for Statistical Computing) by adopting a significance alpha level of 5%. Prediction of target site of circulating miRNA(s) of interest was performed using miRWalk 3.0 (26). Functional enrichment of circulating miRNA targeted genes for Gene Ontology (GO) biological process terms and KEGG pathways was performed using the ClusterProfiler Bioconductor package, and a false discovery rate (FDR)< 0.05.

## Results

### Patient and tumor characteristics

Out of 149 patients treated within the NeoALTTO trastuzumab arm, 126 (85%) had evaluable circulating miRNA profile at the time of surgery (study cohort). The median age at breast cancer diagnosis was 48 years (interquartile range, 43-57). Most of the patients had clinical T2 (64%) and  $\geq$ N1 tumors (72%). Almost half (47%) had ER-positive tumors. pCR was observed in 31% of cases; a total of 40 events were reported at a median follow-up of 6.7 years (interquartile range, 6.1-6.9 years). Patients of the study cohort were randomized in a training (n= 54) and testing set (n= 72). No difference in clinico-pathological characteristics was observed between training and testing sets (Table 1), which were similar to the entire study cohort (Supplementary Table S1).

# End-of-treatment circulating miRNA signature associated with EFS

In the training set, 23 circulating miRNAs were significantly associated with EFS by univariate analysis (Supplementary Table S2). For all these circulating miRNAs, a linear relationship between the EFS probability and their expression was found to be appropriate. By combining these 23 miRNAs into multivariate models following all-subset analysis approach (22), a total of 4 promising prognostic signatures were identified (Table 2). Among these 4 signatures, the model including miR-185-5p, miR-146a-5p, miR-22-3p, was selected as the "best" one. By using the regression coefficient of this model fitted on the whole study cohort, the 3-circulating miRNA signature was generated as following: (-0.062\*miR-185-5p expression) +

TABLE 1 Clinico-pathological features of the training and testing sets.

| Age          | Traning set<br>n= 54 |    | Testing set<br>n= 72 |    |  |  |  |
|--------------|----------------------|----|----------------------|----|--|--|--|
|              | n                    | %  | n                    | %  |  |  |  |
| <50 years    | 28                   | 52 | 39                   | 54 |  |  |  |
| ≥50 years    | 26                   | 48 | 33                   | 46 |  |  |  |
| ER status    |                      |    |                      |    |  |  |  |
| Negative     | 27                   | 50 | 40                   | 56 |  |  |  |
| Positive     | 27                   | 50 | 32                   | 44 |  |  |  |
| Nodal status |                      |    |                      |    |  |  |  |
| N0           | 15                   | 28 | 20                   | 28 |  |  |  |
| ≥N1          | 39                   | 72 | 52                   | 72 |  |  |  |
| Tumor size   |                      |    |                      |    |  |  |  |
| ≤5 cm        | 35                   | 65 | 46                   | 64 |  |  |  |
| >5 cm        | 19                   | 35 | 26                   | 36 |  |  |  |
| pCR          |                      |    |                      |    |  |  |  |
| No           | 37                   | 69 | 50                   | 69 |  |  |  |
| Yes          | 17                   | 31 | 22                   | 31 |  |  |  |
| #Event       | 17                   | 31 | 23                   | 32 |  |  |  |

ER, estrogen receptor; N, clinical nodal status at baseline; pCR, pathological Complete Response; event (i.e. breast cancer relapse after surgery, second primary malignancy, patient death or failure to complete neoadjuvant therapy because of disease progression).

(-0.274\*miR-146a-5p expression)+(0.105\*miR-22-3p expression). Supplementary Figure 2 reports the EFS probability pattern of the 3-circulating miRNA signature. The results of multivariate Cox regression model including these circulating miRNAs are reported in Supplementary Table S3. Notably in the study cohort, the C-statistic of the 3-circulating miRNA signature was 0.62 (95%CI 0.53-0.71), against a C-statistic of 0.58 (95%CI 0.48-0.67) of the model including ER expression, nodal status, tumor size, age and pCR.

## Circulating miRNA signature and clinicopathological variables

Noteworthy, this 3-circulating miRNA signature was evenly distributed among clinico-pathological variables (Figure 1). Moreover, the 3-circulating miRNA signature retained its prognostic performance with respect to EFS even after adjusting for each of the considered clinico-pathological variables (Supplementary Table S4).

# Prognostic value of circulating miRNAs by pCR status

As pCR is a driver of EFS in the NeoALTTO and other neoadjuvant studies, we next analyzed the prognostic value of miRNA according to pCR status. For this purpose, the expression levels of each of the 132 selected miRNAs detected in the study cohort was evaluated thought a multivariate Cox regression model with pCR status and the first-order interaction between miRNA and pCR. We found circulating miR-194-5p with a statistically significant interaction term at alpha level of 0.01 (p.interaction =0.005). The HR estimate in patients with pCR was 1.83 (95%CI 1.14-2.95), and 0.87 (95%CI 0.69-1.10) in those without pCR. Figure 2 reports the EFS probability plot for circulating miR-194-5p expression levels (Log2 RQ) according to pCR status.

# End-of-treatment microRNA added prognostic value

For exploratory purposes, an analysis was performed by adding the interaction terms together with the corresponding main effects (circulating miR-194-5p and pCR) to the 3-circulating miRNA signature. The highest prognostic performance in terms of Cstatistics was observed for the complete multivariate model (Table 3).

# Discussion

Not all early-stage breast cancer patients with residual disease after neoadjuvant therapy have a poor prognosis; on the other hand, some patients achieving a pCR eventually relapse. Therefore, biomarkers are needed to properly identify patients at risk who are ideal candidates for additional post-surgical therapies. Using highthroughput analysis of plasma samples collected within a prospective randomized trial (16), we herein reported the first study investigating circulating microRNAs extensively, and not according to a prespecified candidate panel, at the end of neoadjuvant therapy, and in association with prognosis. Several key findings with biological relevance and clinical potential were identified.

TABLE 2 Prognostic performance of the promising models.

| Promising models                   | Training set*        | Testing set*         |  |
|------------------------------------|----------------------|----------------------|--|
|                                    | C-statistic (95% Cl) | C-statistic (95% CI) |  |
| miR-185-5p, miR-146a-5p, miR-22-3p | 0.705 (0.517; 0.894) | 0.700 (0.519; 0.880) |  |
| miR-146a-5p, miR-15b-3p, miR-22-3p | 0.688 (0.551; 0.825) | 0.698 (0.534; 0.861) |  |
| miR-24-3p, miR-15b-3p, miR-22-3p   | 0.681 (0.541; 0.820) | 0.697 (0.512; 0.882) |  |
| miR-185-5p, miR-23a-3p, miR-22-3p  | 0.683 (0.509; 0.857) | 0.672 (0.520; 0.824) |  |

\*A penalized Cox regression model was implements according to the EPV in the training and testing set.



Firstly, we identified a circulating signature composed of miR-185-5p, miR-146a-5p and miR-22-3p able to discriminate among patients treated with trastuzumab-based therapy with different prognosis. The functions of miRNAs composing our prognostic signature have been associated with tumor-related processes (proliferation, apoptosis, migration/invasion), and response to treatment. Specifically, miR-185-5p seems to act as a tumor suppressor in cancer progression and spreading, at least in central nervous system (27), and gastrointestinal malignancies (28). The precise effects and detailed mechanisms of miR-185-5p in breast cancer are yet to be defined. However, a recently reported pre-clinical study shows that over-expression of miR-185-5p is associated with reduced chemosensitivity (29). Iorio et al. first reported that miR145-5p acts as a tumor suppressor in a variety of tumors, including breast cancer (30); subsequently, miR-145-5p was shown to modulate immune response by targeting the 3'-untranslated region of Tolllike receptor 4 (31), and to increase epithelial-mesenchymal transition through the control of N-cadherin, vimentin and E-cadherin protein expression levels (32). Finally, miR-22 was found both as a tumor suppressor and a promoter in previous studies (33, 34). However, its serum expression levels have already been associated with poor prognosis in breast cancer patients (35). Consistent with these literature data, our analysis showed that both miR-185-5p and miR-146a-5p have a protective prognostic effect, as opposed to miR-22. Furthermore, an integrated analysis of miRNA target gene networks drew much attention because there are many common signaling pathways modulated by differentially expressed mRNAs and shared through GO and KEGG analysis (Supplementary Figure S3). Although circulating miRNAs are not necessarily expressed at the tumor tissue level, it is worthy to note that among the enriched terms, those related to growth factor signaling, metastatic spreading processes and immune response were shared by all dysregulated



#### FIGURE 2

Seven-year Event Free Survival (EFS) probability curves for circulating miR-194-5p values according to pathological complete response (pCR) status. The curve depicts the predicted EFS probability of the circulating miR-194-5p expression levels considered on its continuous scale in patients (A) with pCR and (B) without pCR.

05

### TABLE 3 Prognostic performance of multivariate Cox regression models in the study cohort.

| Model  | C-statistic | 95% Cl |      |
|--|-------------|--------|------|
| 3-circulating miRNA signature + pCR (Yes vs No) <sup>a</sup>                     | 0.62        | 0.53   | 0.71 |
| miR-194-5p + pCR+ (miR-194-5p*pCR) <sup>a</sup>                                  | 0.62        | 0.54   | 0.71 |
| 3-circulating miRNA signature + pCR + miR-194-5p + (miR-194-5p*pCR) <sup>a</sup> | 0.67        | 0.58   | 0.76 |

<sup>a</sup>30-week landmark analysis was performed when pCR was considered in the multivariate Cox regression model; CI, confidence interval.

circulating miRNAs after trastuzumab treatment. These findings are intriguing because development of predictors of recurrence after neoadjuvant therapy is still in its infancy, and presently the only established prognostic factors are stage and hormone receptor status (36). An attempt has been made with TILs (37). However, TILs assessment may be difficult after neoadjuvant therapy (38), so that Asano et al. proposed to combine their evaluation with residual cancer burden, which takes into account tumor dimension, cellularity of tumor bed, and axillary nodal burden (39). In addition TILs data in HER2-positive breast cancer are controversial (40), with several studies suggesting that high TILs values in residual disease after neoadjuvant therapy are associated with worse rather than improved prognosis (reviewed in 37).

Secondly, the 3 circulating miRNA signature ensures a C-statistic of 0.62 (0.53-0.7), instead of 0.58 (0.48-0.67) of the model including clinico-pathological variables, age, stage, estrogen receptor status and pCR. Thus, only three circulating markers assessed at a single time point offer a similar if not superior discriminatory capability of different variables assessed on patient, and primary tumor at baseline or after treatment. Furthermore, the discriminatory capability of the 3-circulating miRNA signature remained significant when clinico-pathological variables were included in multivariate analysis. All these findings support the development of our 3 circulating miRNA signature as a parsimonious and independent prognostic tool.

Thirdly, despite the favorable prognostic effect of invasive disease eradication, a few of patients with pCR eventually relapse (9, 10). Tumor recurrences are in the range of 10-15% at 5 years from surgery, as reported in the large pooled analysis of German Breast Group neoadjuvant studies on 2188 patients (41), and from a recent metaanalysis on 5748 patients (42). These findings are counterintuitive since complete eradication of cancer cells in breast and axilla has been proposed as a maximum effect reflecting the eradication of micrometastatic disease with no room for improvement. Yet, risk of relapse in patients with pCR increases with advanced stage at initial diagnosis, HER2-overexpression, younger age, and premenopausal status. Besides, patients who obtain pCR with the addition of trastuzumab have a better prognosis than patients who obtain pCR with chemotherapy alone (43). Recently, accumulating data suggest that patients who achieve pCR with HER2-dual blockade have a better prognosis than those who achieve pCR with single anti-HER2 agent (44). Although we cannot exclude that these results are due to the imbalance in the size of the subgroups, with fewer patients in the control arm achieving a pCR, these data challenge the idea that all pathological responses born equal and reinforce that of pCR quality. In this context, our data demonstrate for the first time that increased levels of circulating -miR-194-5p are associated with dismal prognosis exclusively in patients achieving a pCR following trastuzumab. miR-194-5p is a p53-responsive miRNA capable of inducing cell cycle arrest and inhibiting cell proliferation, migration, invasion and colony formation (45). These data are inconsistent with our results. However, it has recently been reported that the function miR-194 turns tumorigenic when treatment-sensitive cells promote stem cell survival during the so called "dying for surviving" phenomenon (46). Specifically, miR-194-5p is contained in the exosomes of dying cells and is released to residual ALDH- positive tumor-repopulating cells for their recovery. This finding is consistent with the enrichment of cells with stemness phenotype in occult metastatic lesion (47), and the overexpression of stemness signatures in triple negative breast cancer primary tumor of patients who eventually relapse regardless a pCR (10).

This study has some limitations. First, due to its retrospective nature, biological data were available in most but not all patients included in the NeoALTTO trastuzumab arm. Secondly, the analyzed sample size and the number of breast cancer events prevented subgroup analysis. Finally, the number of patients included in the analysis limits the impact of our findings. Therefore, the results of this study need a further confirmation in a much larger patient population, and with an extended follow-up period. Lastly, the prognostic value of TILs was not evaluated in this study. This would be interesting to be assessed in future studies. In addition, studied patients received additional adjuvant chemo-, ± endocrineand trastuzumab-therapy and we are unaware of the impact of these treatments on miRNAs and vice versa. In view of developing a clinically usable assay, the identification of a limited number of reference miRNAs as well as assay-oriented step(s) should be considered, as described by Verderio et al. (22). As regards the selection of reference miRNAs, they should be properly chosen to resemble the overall mean (19), for example using the procedure we developed (48). The definition of operative procedures for miRs processing and detection as well as the evaluation of the assay performance and robustness, together with the clinical interpretability represent key aspects that should be opportunely addreesd during the assay-oriented step(s).

In conclusion, we have identified a 3 circulating miRNA signature able to differentiate among patients with distinct prognosis after trastuzumab-based neoadjuvant therapy, and the unique circulating-miR-194-5p associated with recurrence after pCR attainment, which warrant further investigation in additional studies. If confirmed, these miRNAs and their possible mechanisms of action could aid the development of new post-neoadjuvant strategies for high risk breast cancer patients. Furthermore, given the absence of tumor tissue in patients attaining a pCR, our results reinforce liquid biopsy as a promising tool to quantify and analyze residual breast cancer burden beyond pathological findings and to predict tumor evolution for post-operative therapy personalization.

## Data availability statement

The raw data supporting the conclusions of this article will be made available by the authors upon reasonable request.

## **Ethics statement**

The studies involving human participants were reviewed and approved by Ethics Committee of Fondazione IRCCS Istituto Nazionale dei Tumori. The patients/participants provided their written informed consent to participate in this study.

# Author contributions

SDC conceived of the presented idea. SDC, CMC, SP and PV contributed to the design and implementation of the research, to the analysis of the results and to the writing of the manuscript. VC and MS verified the analytical methods. All authors contributed to the article and approved the submitted version.

## Funding

The NeoALTTO study was sponsored by GlaxoSmithKline; Lapatinib is an asset of Novartis AG as of March 2, 2015. This substudy was supported by the Italian Ministry of Health to SC. No grant number is applicable, funds were obtained through a law that allows tax-payers to allocate the  $5 \times 1000$  share of their payments to research.

# Acknowledgments

These data were partially reported as oral presentation at Società Italiana di Cancerologia (SIC) Conference, Virtual Event, October 27-28, 2021.

# References

1. Korde LA, Somerfield MR, Carey LA, Crews JR, Denduluri N, Hwang ES, et al. Neoadjuvant chemotherapy, endocrine therapy, and targeted therapy for breast cancer: ASCO guideline. *J Clin Oncol* (2021) 39:1485–505. doi: 10.1200/JCO.20.03399

2. Buchholz TA, Mittendorf EA, Hunt KK. Surgical considerations after neoadjuvant chemotherapy: Breast conservation therapy. J Natl Cancer Inst Monogr (2015) 51:11–4. doi: 10.1093/jncimonographs/lgv020

3. Classe JM, Loaec C, Gimbergues P, Alran S, de Lara CT, Dupre PF, et al. Sentinel lymph node biopsy without axillary lymphadenectomy after neoadjuvant chemotherapy is accurate and safe for selected patients: the GANEA 2 study. *Breast Cancer Res Treat* (2019) 173:343–52. doi: 10.1007/s10549-018-5004-7

4. Cortazar P, Zhang L, Untch M, Mehta K, Costantino JP, Wolmark N, et al. Pathological complete response and long-term clinical benefit in breast cancer: The CTNeoBC pooled analysis. *Lancet* (2014) 384:164–72. doi: 10.1016/S0140-6736(13)62422-8

5. Broglio KR, Quintana M, Foster M, Olinger M, McGlothlin A, Berry SM, et al. Association of pathologic complete response to neoadjuvant therapy in HER2-positive

# **Conflict of interest**

SDC received fees for medical education from Novartis, Pierre-Fabre, and IQVIA; and the research grant IG 20774 from Fondazione Associazione Italiana Ricerca contro il Cancro AIRC; and served as "ad hoc" medical advisor for Medica Scientia Innovation Research MEDSIR, Barcelona Spain. SE-A received grants from Novartis during the conduct of the ALTTO study, and grants from Roche/Genentech and Pfizer outside of the submitted work. MI is employed at Novartis Pharma. HG received honoraria as speaker from AstraZeneca, Roche, BMS, and Novartis. PN received grants from Novartis, Roche/Genentech, MSD Oncology, Bayer, and Targos outside the submitted work. MB's institution received a research grant for the conduct of the NeoALTTO study. JH received advisory boards and honoraria from Roche and research grants from Novartis. DC received grant support, consulting fees, and fees for independent data monitoring committee work from Roche, grant support and consulting fees from Novartis, fees for independent data monitoring committee work from Synthon, and consulting fees from Daiichi Sankyo, Samsung BIOEPSIS, Puma Biotechnology, Seattle Genetics, and Zymeworks all support paid to his institution.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

# Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

# Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fonc.2022.1028825/ full#supplementary-material

breast cancer with long-term outcomes: A meta-analysis. JAMA Oncol (2016) 2:751–60. doi: 10.1001/jamaoncol.2015.6113

6. Available at: https://clinicaltrials.gov (Accessed August 17, 2022).

7. von Minckwitz G, Huang CS, Mano MS, Loibl S, Mamounas EP, Untch M, et al. Trastuzumab emtansine for residual invasive HER2-positive breast cancer. *N Engl J Med* (2019) 380:617–28. doi: 10.1056/NEJMoa1814017

8. Huober J, Holmes E, Baselga J, de Azambuja E, Untch M, Fumagalli D, et al. Survival outcomes of the NeoALTTO study (BIG 1-06): updated results of a randomised multicenter phase III neoadjuvant clinical trial in patients with HER2-positive primary breast cancer. *Eur J Cancer* (2019) 118:169–77. doi: 10.1016/j.ejca.2019.04.038

9. Xie LY, Wang K, Chen HL, Shi YX, Zhang YQ, Lin HY, et al. Markers associated with tumor recurrence in patients with breast cancer achieving a pathologic complete response after neoadjuvant chemotherapy. *Front Oncol* (2022) 12:860475. doi: 10.3389/ fonc.2022.860475

10. Bruzas S, Gluz O, Harbeck N, Schmid P, Cortes J, Blohmer J, et al. Gene signatures in patients with early breast cancer and relapse despite pathologic complete response. *NPJ Breast Cancer* (2022) 8:42. doi: 10.1038/s41523-022-00403-3

11. Afzal S, Hassan M, Ullah S, Abbas H, Tawakkal F, Khan MA. Breast cancer; discovery of novel diagnostic biomarkers, drug resistance, and therapeutic implication. *Front Mol Biosci* (2022) 9:783450. doi: 10.3389/fmolb.2022.783450

12. Müller V, Gade S, Steinbach B, Loibl S, von Minckwitz G, Untch M, et al. Changes in serum levels of miR-21, miR-210, and miR-373 in HER2-positive breast cancer patients undergoing neoadjuvant therapy: a translational research project within the geparquinto trial. *Breast Cancer Res Treat* (2014) 147:61–8. doi: 10.1007/s10549-014-3079-3

13. Du F, Yuan P, Zhao ZT, Yang Z, Wang T, Zhao JD, et al. A miRNA-based signature predicts development of disease recurrence in HER2 positive breast cancer after adjuvant trastuzumab-based treatment. *Sci Rep* (2016) 6:33825. doi: 10.1038/srep33825

14. Jung EJ, Santarpia L, Kim J, Esteva FJ, Moretti E, Buzdar AU, et al. Plasma microRNA 210 levels correlate with sensitivity to trastuzumab and tumor presence in breast cancer patients. *Cancer* (2012) 118:2603–14. doi: 10.1002/cncr.26565

15. Di Cosimo S, Appierto V, Pizzamiglio S, Tiberio P, Iorio MV, Hilbers F, et al. Plasma miRNA levels for predicting therapeutic response to neoadjuvant treatment in HER2-positive breast cancer: Results from the NeoALTTO trial. *Clin Cancer Res* (2019) 25:3887–95. doi: 10.1158/1078-0432.CCR-18-2507

16. Baselga J, Bradbury I, Eidtmann H, Di Cosimo S, de Azambuja E, Aura C, et al. Lapatinib with trastuzumab for HER2-positive early breast cancer (NeoALTTO): A randomised, open-label, multicentre, phase 3 trial. *Lancet* (2012) 379:633–40. doi: 10.1016/S0140-6736(11)61847-3

17. Blondal T, Jensby Nielsen S, Baker A, Andreasen D, Mouritzen P, Wrang Teilum M, et al. Assessing sample and miRNA profile quality in serum and plasma or other biofluids. *Methods* (2013) 59:S1–6. doi: 10.1016/j.ymeth.2012.09.015

18. Livak KJ and Schmittgen TD. Analysis of relative gene expression data using real-time quantitative PCR and the 2- $\Delta\Delta$ Ct method. *Methods* (2001) 25:402–8. doi: 10.1006/meth.2001.1262

19. Mestdagh P, Hartmann N, Baeriswyl L, Andreasen D, Bernard N, Chen C, et al. Evaluation of quantitative miRNA expression platforms in the microRNA quality control (miRQC) study. *Nat Methods* (2014) 11:809–15. doi: 10.1038/nmeth.3014

20. Durrleman S, Simon R. Flexible regression models with cubic splines. *Stat Med* (1989) 8:551-61. doi: 10.1002/sim.4780080504

21. Concato J, Peduzzi P, Holford TR, Feinstein AR. Importance of events per independent variable in proportional hazards analysis. i. background, goals, and general strategy. *J Clin Epidemiol* (1995) 48:1495–501. doi: 10.1016/0895-4356(95) 00510-2

22. Verderio P, Bottelli S, Pizzamiglio S, Ciniselli CM. Developing miRNA signatures: A multivariate prospective. *Br J Cancer* (2016) 115:1–4. doi: 10.1038/bjc.2016.164

23. Heinze G, Schemper M. A solution to the problem of monotone likelihood in cox regression. *Biometrics* (2001) 57:114–9. doi: 10.1111/j.0006-341X.2001.00114.x

24. Uno H, Cai T, Pencina MJ, D'Agostino RB, Wei LJ. On the c-statistics for evaluating overall adequacy of risk prediction procedures with censored survival data. *Stat Med* (2011) 30:1105–17. doi: 10.1002/sim.4154

25. Kirschner MB, Edelman JJB, Kao SCH, Vallely MP, van Zandwijk N, Reid G. The impact of hemolysis on cell-free microRNA biomarkers. *Front Genet* (2013) 4:94. doi: 10.3389/fgene.2013.00094

26. Sticht C, de la Torre C, Parveen A, Gretz N. miRWalk: An online resource for prediction of microRNA binding sites. *PLos One* (2018) 13:e0206239. doi: 10.1371/journal.pone.0206239

27. Wu W, Yu T, Wu Y, Tian W, Zhang J, Wang Y. The miR155HG/miR-185/ANXA2 loop contributes to glioblastoma growth and progression. *J Exp Clin Cancer Res* (2019) 38:133. doi: 10.1186/s13046-019-1132-0

28. Yuan M, Zhang X, Zhang J, Wang K, Zhang Y, Shang W, et al. DC-SIGN-LEF1/ TCF1-miR-185 feedback loop promotes colorectal cancer invasion and metastasis. *Cell Death Differ* (2020) 27:379–95. doi: 10.1038/s41418-019-0361-2

29. Luo I., Zhang J, Tang H, Zhai D, Huang D, Ling L, et al. LncRNA SNORD3A specifically sensitizes breast cancer cells to 5-FU by sponging miR-185-5p to enhance UMPS expression. *Cell Death Dis* (2020) 11:329. doi: 10.1038/s41419-020-2557-2

30. Iorio MV, Ferracin M, Liu CG, Veronese A, Spizzo R, Sabbioni S, et al. MicroRNA gene expression deregulation in human breast cancer. *Cancer Res* (2005) 65:7065–70. doi: 10.1158/0008-5472.CAN-05-1783

31. Jin C, Wang A, Liu L, Wang G, Li G, Han Z. miR-145-5p inhibits tumor occurrence and metastasis through the NF-κB signaling pathway by targeting TLR4 in malignant melanoma. *J Cell Biochem* (2019) 120:11115–26. doi: 10.1002/jcb.28388

32. Liu Q, Chen J, Wang B, Zheng Y, Wan Y, Wang Y, et al. miR-145 modulates epithelial-mesenchymal transition and invasion by targeting ZEB2 in non-small cell lung cancer cell lines. *J Cell Biochem* (2019) 120:8409–18. doi: 10.1002/jcb.28126

33. Yang F, Hu Y, liu HX, Wan YJ. MiR-22-silenced cyclin a expression in colon and liver cancer cells is regulated by bile acid receptor. *J Biol Chem* (2015) 290:6507–15. doi: 10.1074/jbc.M114.620369

34. Lee JH, Park SJ, Jeong SY, Kim MJ, Jun S, Lee HS. MicroRNA-22 suppresses DNA repair and promotes genomic instability through targeting of MDC1. *Cancer Res* (2015) 75:1298–310. doi: 10.1158/0008-5472.can-14-2783

35. Shao Y, Yao Y, Xiao P, Yang X, Zhang D. Serum miR-22 could be a potential biomarker for the prognosis of breast cancer. *Clin Lab* (2019) 65:4. doi: 10.7754/ clin.lab.2018.180825

36. Guarneri V, Broglio K, Kau SW, Cristofanilli M, Buzdar AU, Valero V, et al. Prognostic value of pathologic complete response after primary chemotherapy in relation to hormone receptor status and other factors. *J Clin Oncol* (2006) 24:1037–44. doi: 10.1200/JCO.2005.02.6914

37. Pelizzari G, Gerratana L, Basile D, Fanotto V, Bartoletti M, Liguori A, et al. Postneoadjuvant strategies in breast cancer: From risk assessment to treatment escalation. *Cancer Treat Rev* (2019) 72:7–14. doi: 10.1016/j.ctrv.2018.10.014

38. Dieci MV, Radosevic-Robin N, Fineberg S, van den Eynden G, Ternes N, Penault-Llorca F, et al. Update on tumor-infiltrating lymphocytes (TILs) in breast cancer, including recommendations to assess TILs in residual disease after neoadjuvant therapy and in carcinoma in situ: A report of the international immuno-oncology biomarker working group on breast cancer. *Semin Cancer Biol* (2018) 52:16–25. doi: 10.1016/ j.semcancer.2017.10.003

39. Asano Y, Kashiwagi S, Goto W, Takada K, Takahashi K, Hatano T, et al. Prediction of survival after neoadjuvant chemotherapy for breast cancer by evaluation of tumor-infiltrating lymphocytes and residual cancer burden. *BMC Cancer* (2017) 17:888. doi: 10.1186/s12885-017-3927-8

40. Hamy AS, Pierga JY, Sabaila A, Laas E, Bonsang-Kitzis H, Laurents C, et al. Stromal lymphocyte infiltration after neoadjuvant chemotherapy is associated with aggressive residual disease and lower disease-free survival in HER2-positive breast cancer. *Ann Oncol Off J Eur Soc Med Oncol* (2017) 28:2233–40. doi: 10.1093/annonc/mdx309

41. Huober J, Schneeweiss A, Blohmer JU, Denkert C, Tesch H, Hanusch CA, et al. Factors predicting relapse in early breast cancer patients with a pathological complete response after neoadjuvant therapy-results of a pooled analysis based on the GBG metadatabase. *Cancer Res* (2019) 79:P2–08-01. doi: 10.1158/1538-7445.SABCS18-P2-08-01

42. Spring LM, Fell G, Arfe A, Sharma C, Greenup R, Reynolds KL, et al. Pathologic complete response after neoadjuvant chemotherapy and impact on breast cancer recurrence and survival: A comprehensive meta-analysis. *Clin Cancer Res* (2020) 26:2838–48. doi: 10.1158/1078-0432.CCR-19-3492

43. Gianni L, Eiermann W, Semiglazov V, Lluch A, Tjulandin S, Zambetti M, et al. Neoadjuvant and adjuvant trastuzumab in patients with HER2-positive locally advanced breast cancer (NOAH): Follow-up of a randomised controlled superiority trial with a parallel HER2-negative cohort. *Lancet Oncol* (2014) 15:640–7. doi: 10.1016/S1470-2045 (14)70080-4

44. Swain SM, Macharia H, Cortes J, Dang C, Gianni L, Hurvitz S, et al. Risk of recurrence and death in patients with early HER2-positive breast cancer who achieve a pathological complete response after different types of HER2-targeted therapy: A retrospective exploratory analysis. 2019 San Antonio Breast Cancer Symposium. Abstract P1-18-01. *Cancer Res* (2020) 80(4\_Supplement):P1-18-01.

45. Kurata JS, Lin RJ. MicroRNA-focused CRISPR-Cas9 library screen reveals fitnessassociated miRNAs. RNA (2018) 24:966–81. doi: 10.1261/rna.066282.118

46. Obenauf AC, Zou Y, Ji AL, Vanharanta S, Shu W, Shi H, et al. Therapy-induced tumour secretomes promote resistance and tumour progression. *Nature* (2015) 520:368.72. doi: 10.1038/nature14336

47. Gooding AJ, Schiemann WP. Epithelial-mesenchymal transition programs and cancer stem cell phenotypes: Mediators of breast cancer therapy resistance. *Mol Cancer Res* (2020) 18:1257–70. doi: 10.1158/1541-7786.MCR-20-0067

48. Verderio P, Bottelli S, Ciniselli CM, Pierotti MA, Gariboldi M, Pizzamiglio S. NqA: An r-based algorithm for the normalization and analysis of microRNA quantitative realtime polymerase chain reaction data. *Anal Biochem* (2014) 46:7–9. doi: 10.1016/ j.ab.2014.05.020